



## MyD88 Signaling Inhibits Protective Immunity to the Gastrointestinal Helminth Parasite *Heligmosomoides polygyrus*

This information is current as of August 30, 2019.

Lisa A. Reynolds, Yvonne H Marcus, Katherine A. Smith, Lauren M. Webb, James P. Hewitson, Ewan A. Ross, Sheila Brown, Satoshi Uematsu, Shizuo Akira, David Gray, Mohini Gray, Andrew S. MacDonald, Adam F. Cunningham and Rick M. Maizels

*J Immunol* 2014; 193:2984-2993; Prepublished online 11 August 2014;

doi: 10.4049/jimmunol.1401056

<http://www.jimmunol.org/content/193/6/2984>

**Supplementary Material** <http://www.jimmunol.org/content/suppl/2014/08/09/jimmunol.1401056.DCSupplemental>

**References** This article **cites 69 articles**, 24 of which you can access for free at: <http://www.jimmunol.org/content/193/6/2984.full#ref-list-1>

**Why *The JI*? Submit online.**

- **Rapid Reviews! 30 days\*** from submission to initial decision
- **No Triage!** Every submission reviewed by practicing scientists
- **Fast Publication!** 4 weeks from acceptance to publication

*\*average*

**Subscription** Information about subscribing to *The Journal of Immunology* is online at: <http://jimmunol.org/subscription>

**Permissions** Submit copyright permission requests at: <http://www.aai.org/About/Publications/JI/copyright.html>

**Email Alerts** Receive free email-alerts when new articles cite this article. Sign up at: <http://jimmunol.org/alerts>



# MyD88 Signaling Inhibits Protective Immunity to the Gastrointestinal Helminth Parasite *Heligmosomoides polygyrus*

Lisa A. Reynolds,<sup>\*,†,1</sup> Yvonne Harcus,<sup>\*,†</sup> Katherine A. Smith,<sup>\*,†,2</sup> Lauren M. Webb,<sup>\*,†,3</sup> James P. Hewitson,<sup>\*,†,4</sup> Ewan A. Ross,<sup>‡</sup> Sheila Brown,<sup>\*,†,3</sup> Satoshi Uematsu,<sup>§</sup> Shizuo Akira,<sup>¶,||</sup> David Gray,<sup>\*,†</sup> Mohini Gray,<sup>\*,†</sup> Andrew S. MacDonald,<sup>\*,†,3</sup> Adam F. Cunningham,<sup>‡</sup> and Rick M. Maizels<sup>\*,†</sup>

Helminth parasites remain one of the most common causes of infections worldwide, yet little is still known about the immune signaling pathways that control their expulsion. C57BL/6 mice are chronically susceptible to infection with the gastrointestinal helminth parasite *Heligmosomoides polygyrus*. In this article, we report that C57BL/6 mice lacking the adapter protein MyD88, which mediates signaling by TLRs and IL-1 family members, showed enhanced immunity to *H. polygyrus* infection. Alongside increased parasite expulsion, MyD88-deficient mice showed heightened IL-4 and IL-17A production from mesenteric lymph node CD4<sup>+</sup> cells. In addition, MyD88<sup>-/-</sup> mice developed substantial numbers of intestinal granulomas around the site of infection, which were not seen in MyD88-sufficient C57BL/6 mice, nor when signaling through the adapter protein TRIF (TIR domain-containing adapter-inducing IFN- $\beta$  adapter protein) was also ablated. Mice deficient solely in TLR2, TLR4, TLR5, or TLR9 did not show enhanced parasite expulsion, suggesting that these TLRs signal redundantly to maintain *H. polygyrus* susceptibility in wild-type mice. To further investigate signaling pathways that are MyD88 dependent, we infected IL-1R1<sup>-/-</sup> mice with *H. polygyrus*. This genotype displayed heightened granuloma numbers compared with wild-type mice, but without increased parasite expulsion. Thus, the IL-1R–MyD88 pathway is implicated in inhibiting granuloma formation; however, protective immunity in MyD88-deficient mice appears to be granuloma independent. Like IL-1R1<sup>-/-</sup> and MyD88<sup>-/-</sup> mice, animals lacking signaling through the type 1 IFN receptor (i.e., IFNAR1<sup>-/-</sup>) also developed intestinal granulomas. Hence, IL-1R1, MyD88, and type 1 IFN receptor signaling may provide pathways to impede granuloma formation in vivo, but additional MyD88-mediated signals are associated with inhibition of protective immunity in susceptible C57BL/6 mice. *The Journal of Immunology*, 2014, 193: 2984–2993.

Intestinal helminth parasites are highly prevalent worldwide (1), yet little is understood of the signaling pathways that lead to their immune exclusion. *Heligmosomoides polygyrus* is a natural intestinal nematode parasite of mice that can be maintained in the laboratory. Adult worms reside in the small intestine, alongside commensal organisms and dietary Ags, and release eggs into the feces for onward transmission. This tractable model system has been widely used to study factors affecting susceptibility to helminth infection (2–5).

Following oral ingestion of *H. polygyrus* larvae, the small intestinal epithelial cell barrier is disrupted first as the parasite's infective larvae enter the small intestinal submucosa (by 24 h post infection), and later as the adult worms emerge to take up residence in the intestinal lumen (by 10 d post infection) (4). In most strains of mice, *H. polygyrus* establishes a long-term chronic in-

fection, associated with the expansion of regulatory T cells (Tregs) (6–10), which inhibit host effector responses. However, some mouse strains express immunity to primary infection, as measured by diminished release of eggs in the feces, and faster expulsion of adult worms between 14 and 28 d of infection (10–12). Immunity can also be generated in most genotypes by prior immunization with parasite-secreted Ags (13, 14) or by abbreviating primary infection through drug treatment (15, 16).

Around the submucosal sites of larval invasion, type 2 granulomas consisting primarily of macrophages and neutrophils form (16–18). The number of intestinal granulomas generally correlates with the resistance phenotype of mouse strains, with the most resistant genotypes developing greater granuloma numbers following infection, which persist well after adult worms have emerged into the intestinal lumen, and have even been expelled

\*Institute of Immunology and Infection Research, University of Edinburgh, Edinburgh, EH9 3JT, United Kingdom; <sup>†</sup>Centre for Immunity, Infection and Evolution, University of Edinburgh, Edinburgh, EH9 3JT, United Kingdom; <sup>‡</sup>Medical Research Council Centre for Immune Regulation, Institute of Microbiology and Infection, School of Immunity and Infection, University of Birmingham, Birmingham, B15 2TT, United Kingdom; <sup>§</sup>Division of Innate Immune Regulation, International Research and Development Center for Mucosal Vaccines, Institute of Medical Science, The University of Tokyo, Shirokanedai, Minato-ku, Tokyo, 108-8639, Japan; <sup>¶</sup>Department of Host Defense, Research Institute for Microbial Diseases, Osaka University, Suita, Osaka 565-0871, Japan; and <sup>||</sup>Laboratory of Host Defense, World Premier Institute Immunology Frontier Research Center, Osaka University, Suita, Osaka 565-0871, Japan

<sup>1</sup>Current address: Michael Smith Laboratories, University of British Columbia, Vancouver, BC, Canada.

<sup>2</sup>Current address: Cardiff Institute of Infection & Immunity, Cardiff, U.K.

<sup>3</sup>Current address: Manchester Collaborative Centre for Inflammation Research, University of Manchester, Manchester, U.K.

<sup>4</sup>Current address: Centre for Immunology and Infection, University of York, York, U.K.

Received for publication April 24, 2014. Accepted for publication July 15, 2014.

This work was supported by Wellcome Trust Grants Ref. 090281 (to R.M.M.) and Ref. 086629 (to L.A.R.) and by Medical Research Council Grant G0701437 (to L.M.W. and A.S.M.).

Address correspondence and reprint requests to Dr. Rick M. Maizels, Institute of Immunology and Infection Research, University of Edinburgh, Edinburgh, EH9 3JT, U.K. E-mail address: r.maizels@ed.ac.uk

The online version of this article contains supplemental material.

Abbreviations used in this article: IFNAR, type 1 IFN receptor; MLN, mesenteric lymph node; PRR, pattern recognition receptor; Treg, regulatory T cell; TRIF, TIR domain-containing adapter-inducing IFN- $\beta$  adapter protein.

This is an open-access article distributed under the terms of the [CC-BY 3.0 Unported license](http://creativecommons.org/licenses/by/3.0/).

Copyright © 2014 The Authors 0022-1767/14

(10, 16, 19). However, it is not known whether granuloma formation is necessary or sufficient for immunity to infection.

*H. polygyrus* shares its intestinal niche with many commensal microorganisms, and its penetration of the epithelial barrier may cause aberrant host exposure to microbial stimuli. In this respect, it is interesting to note previous reports that germ-free mice are more resistant to infection with *H. polygyrus* than are conventionally raised mice (20–22), leading us to hypothesize that direct or indirect signals from the intestinal microbiota modulate the outcome of infection.

The immune system recognizes highly conserved bacterial components through pattern recognition receptors (PRRs), which include the TLRs, nucleotide-binding oligomerization domain protein-like receptors, and C-type lectin receptors. Recognition of intestinal bacteria by these PRRs is required to maintain gut homeostasis in conditions in which epithelial cell barrier function is lost. For example, deficiencies in PRR signaling lead to heightened morbidity and mortality after dextran sodium sulfate treatment to experimentally induce colitis (23–25). Following epithelial cell damage, inflammatory and reparative cytokines, including TNF and IL-6, that can mediate repair are induced, and it is the recognition of intestinal microbes that provides the trigger for production of these cytokines (23).

It is likely that during periods of epithelial barrier disruption by *H. polygyrus*, ligation of PRRs by the intestinal microbiota occurs, but how this immune stimulation affects the antiparasite immune response has not yet been considered. A recent study has shown that when the integrity of the gut epithelium is disrupted during *Toxoplasma gondii* infection, a microflora-specific T cell response is mounted (26). Thus, it is reasonable to assume that the presence of the microflora will influence the immune environment surrounding infective *H. polygyrus* larvae.

In this article, we set out to investigate how immunity to *H. polygyrus* was affected when C57BL/6 mice, which typically are chronically susceptible to infection with this parasite, lacked the adapter protein MyD88, a major mediator of signaling through TLRs (27) and IL-1 family members (28). Strikingly, we show that MyD88<sup>-/-</sup> mice were rendered more resistant to *H. polygyrus*. This resistance was associated with a heightened Th2 response, as well as a dramatic increase in the number of granulomas being produced following *H. polygyrus* infection, which rarely formed in C57BL/6 wild-type control mice. We demonstrate that granuloma formation is inhibited by a pathway involving IL-1R1, type 1 IFN signaling, and MyD88, but distinct MyD88-dependent mechanisms maintain susceptibility to helminth infection in wild-type mice.

## Materials and Methods

### Mice

C57BL/6, TLR2<sup>-/-</sup>, TLR4<sup>-/-</sup>, TLR9<sup>-/-</sup>, MyD88<sup>-/-</sup>, TIR domain-containing adapter-inducing IFN- $\beta$  adapter protein (TRIF)<sup>-/-</sup>, type 1 IFN receptor (IFNAR1)<sup>-/-</sup>, and MyD88<sup>-/-</sup> IFNAR1<sup>-/-</sup> mice were bred in house at the University of Edinburgh. IL-1R1<sup>-/-</sup> mice were bred at the University of Oxford, and TLR5<sup>-/-</sup> mice at the University of Birmingham, U.K. All transgenic mice were on a C57BL/6 background. To equalize the microbiota compositions prior to beginning each experiment, bedding was mixed between C57BL/6 and transgenic male mouse cages, and female C57BL/6 and transgenic mice were cohoused for at least 1 wk prior to, and throughout the duration of, each experiment. Mice were housed in individually ventilated cages, and all experiments performed complied with U.K. Home Office guidelines.

### Parasites and granuloma measurements

Mice were infected by oral gavage of 200 *H. polygyrus bakeri* third stage larvae, which were obtained from fecal cultures of *H. polygyrus*-infected mice. *H. polygyrus* egg counts were performed on fecal pellets using a McMaster 2 cell counter (Hawksley) at 2 wk and 4 wk following in-

fection, and 4 wk following infection, intestinal adult worms and granulomas were enumerated. Granulomas were macroscopically counted along the entire length of the small intestinal tract and were not present in the large intestine.

### Cell staining and flow cytometry

Mesenteric lymph nodes (MLNs) were manually dissected, and single-cell suspensions were made by passing cells through a 70  $\mu$ m cell strainer in RPMI 1640 media (Life Technologies) containing 10% FCS (Hyclone), 2 mM L-glutamine (Life Technologies), and 100 U/ml penicillin and 100  $\mu$ g/ml streptomycin (Life Technologies).

Cells were stained directly ex vivo for Foxp3 measurements or, for intracellular cytokine measurements, restimulated with 0.5  $\mu$ g/ml PMA and 1  $\mu$ g/ml ionomycin for 3.5 h, with 10  $\mu$ g/ml brefeldin A included for the final 2.5 h. Cells were stained for 20 min at 4°C with Abs to the surface marker CD4 (RM4-5; BD Pharmingen), and CD103 conjugated to biotin (M290; BD Pharmingen) followed by PerCP-streptavidin (BD Pharmingen). Cells were fixed according to the manufacturer's instructions with Fix/Perm (eBiosciences) for Foxp3 measurements, or Cytofix/CytoPerm (BD) for intracellular cytokine measurements, and then stained for 20 min at 4°C with Abs to Foxp3 (FJK-16s; eBioscience), IL-4 (11B11; BioLegend), IFN- $\gamma$  (XMG1.2; BioLegend), IL-17A (TC11-18H10.1; BioLegend), or the relevant isotype controls. Marker expression was measured on FACSCanto (BD) or LSR II (BD) flow cytometers, and data were analyzed using FlowJo software (TreeStar).

### Statistics

Data were first examined for normality. Where data were normally distributed, and comparisons between two groups were being made, unpaired *t* tests were performed, and where comparisons between multiple groups were being made, one-way ANOVA tests followed by Tukey tests were used. In situations when data were not normally distributed, Mann-Whitney tests were used to compare two groups or, for comparisons between multiple groups, Kruskal-Wallis tests followed by Dunn tests were used.

Average bars shown on graphs indicate the mean value for parametric data, or the median value for nonparametric data. The *p* values are as follows: \**p*  $\leq$  0.05, \*\**p*  $\leq$  0.01, \*\*\**p*  $\leq$  0.001.

Data from multiple experiments were pooled where indicated, and only when no statistical differences existed between separate data sets.

## Results

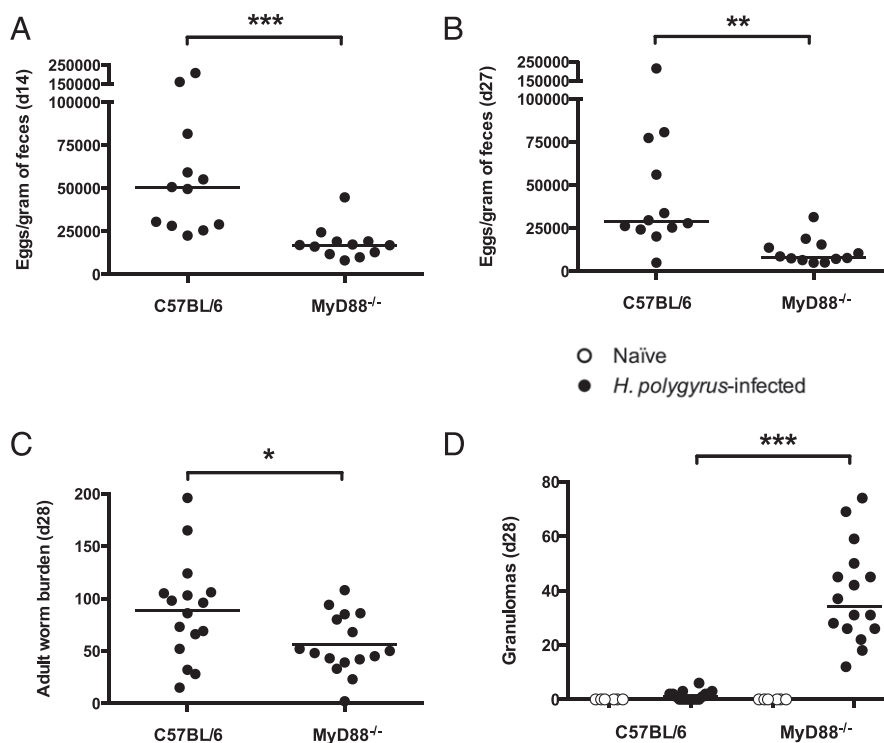
### MyD88 deficiency renders mice more resistant to *H. polygyrus* than wild-type C57BL/6 mice

To investigate whether PRR signaling modulates the outcome of a helminth infection, we first compared mice lacking the adapter protein MyD88, through which many TLRs signal (27), with wild-type C57BL/6 mice for their susceptibility to *H. polygyrus*. Although wild-type C57BL/6 mice are chronically susceptible to this parasite, a lack of MyD88 signaling was found to render mice markedly more resistant to infection, with a lower *H. polygyrus* egg output seen at days 14 and 27 post infection (Fig. 1A, 1B), an indication of lower worm fitness, which correlates with stronger immune responsiveness (10). In addition, significantly fewer adult worms remained in the MyD88-deficient host 28 d following infection (Fig. 1C).

A striking phenotype in MyD88<sup>-/-</sup> mice was the increased number of granulomas that formed along the small intestinal tract following *H. polygyrus* infection, which were rarely observed at this time point in wild-type C57BL/6 mice (Fig. 1D). Granuloma formation has previously been associated with more rapid helminth expulsion, with increased numbers of granulomas evident in more genetically resistant mouse strains (10, 19).

To determine whether MyD88 signaling by hematopoietic or nonhematopoietic cells is important for controlling immunity to *H. polygyrus*, bone marrow chimeras were generated in which either the hematopoietic or the nonhematopoietic compartment lacked MyD88 expression. Neither MyD88 deficiency solely within hematopoietic, nor within nonhematopoietic cells, was sufficient to

**FIGURE 1.** MyD88 deficiency renders mice more resistant to *H. polygyrus* than wild-type C57BL/6 mice. C57BL/6 and MyD88<sup>-/-</sup> mice were left naive or were infected with 200 *H. polygyrus* third stage larvae. **(A)** *H. polygyrus* eggs per gram of feces taken 14 d post infection. **(B)** *H. polygyrus* eggs per gram of feces taken 27 d post infection. **(C)** Adult *H. polygyrus* numbers recovered from the intestinal tract 28 d post infection. **(D)** Number of granulomas formed along the small intestinal tract 28 d post infection. Data shown in (A) and (B) are pooled from two independent experiments, each with four to eight mice per group; data shown in (C) and (D) are pooled from three independent experiments, each with four to eight mice per group. \**p* ≤ 0.05, \*\**p* ≤ 0.01, \*\*\**p* ≤ 0.001.



result in the heightened immunity of mice completely deficient in MyD88, suggesting that MyD88 signaling by both compartments contributes to the susceptibility of C57BL/6 mice to *H. polygyrus* (Supplemental Fig. 1).

*Mice singly deficient in specific TLRs harbor H. polygyrus burdens similar to those in wild-type C57BL/6 mice*

MyD88 is used for signaling by TLRs (27). To address whether the increased resistance to *H. polygyrus* in MyD88<sup>-/-</sup> mice was due to lack of signaling through a single, specific TLR, mice deficient in individual TLRs known to sense bacterial products were assessed for their susceptibility to *H. polygyrus*.

The ability of MyD88-deficient mice to expel *H. polygyrus* more rapidly was not reproduced in the absence of TLR2 (Fig. 2A), TLR4 (Fig. 2B), TLR5 (Fig. 2C), or TLR9 (Fig. 2D) alone, leading to the possibility that TLRs signal redundantly to maintain *H. polygyrus* susceptibility on the C57BL/6 background. However, in TLR2<sup>-/-</sup> mice *H. polygyrus* showed reduced egg output by day 28 post infection, compared with wild-type C57BL/6 controls (Fig. 2E), although considerable variability in egg output was seen in these mice. In contrast, neither TLR4<sup>-/-</sup>, TLR5<sup>-/-</sup>, nor TLR9<sup>-/-</sup> mice showed significant differences in *H. polygyrus* egg output, compared with wild-type controls (Fig. 2F).

*MyD88<sup>-/-</sup> mice mount a stronger CD4<sup>+</sup> T cell IL-4 and IL-17A response following H. polygyrus infection*

We next investigated how T cell responses, which play an important role in mediating immunity to a primary *H. polygyrus* infection (4, 29), differed between wild-type C57BL/6 and MyD88-deficient mice. At 28 d post *H. polygyrus* infection, levels of IL-4 and of IL-17A being produced by CD4<sup>+</sup> T cells in the MLN cells of MyD88-deficient mice were higher than in wild-type C57BL/6 counterparts (Fig. 3A, 3B), whereas IFN- $\gamma$  levels were similar between infected groups (Fig. 3C). IL-4 is recognized as one of the most critical cytokines for mediating protective immunity to *H. polygyrus* (4, 30); however, Ab-mediated depletion of IL-17A has recently been found to have little effect on the outcome of

primary infection with this parasite (31). Despite the reduced *H. polygyrus* egg output in TLR2<sup>-/-</sup> mice compared with wild-type mice (Fig. 2E), MLN CD4<sup>+</sup> T cells from naive and 28-d *H. polygyrus*-infected TLR2<sup>-/-</sup> mice produced levels of IL-4, IL-17A, and IFN- $\gamma$  comparable to those from wild-type mice (Supplemental Fig. 2).

*TLR signaling contributes differentially to Treg proportions following H. polygyrus infection*

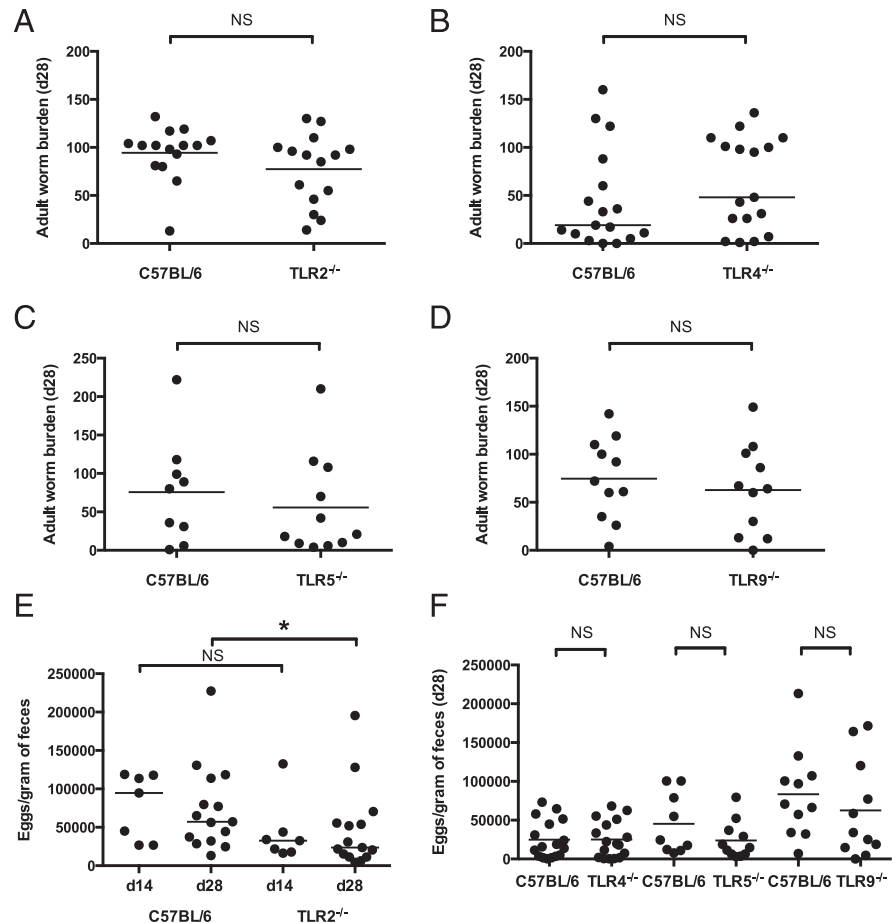
As MyD88-deficient mice produced more signature cytokines of both Th2 and Th17 subsets following *H. polygyrus* infection, we then examined whether the proportions and activation status of Tregs were abnormal in these mice. Surprisingly, no deficiencies in Foxp3<sup>+</sup>CD4<sup>+</sup> T cell proportions, or in their expression of CD103, were seen in the MLNs of MyD88-deficient mice (Fig. 4A, 4B).

Dysregulation of Treg proportions were, however, seen in singly TLR-deficient mice. Both TLR2- and TLR5-deficient mice had a lower proportion of Foxp3<sup>+</sup>CD4<sup>+</sup> MLN T cells following *H. polygyrus* infection than did wild-type C57BL/6 mice (Fig. 4C, 4G). A greater proportion of Foxp3<sup>+</sup>CD4<sup>+</sup> MLN T cells in TLR2-deficient mice expressed CD103 (Fig. 4D) than in wild-type C57BL/6, whereas CD103 expression levels on Foxp3<sup>+</sup>CD4<sup>+</sup> MLN T cells were similar between wild-type C57BL/6 and TLR5-deficient mice (Fig. 4H). Conversely, TLR9-deficient mice had a higher proportion of Foxp3<sup>+</sup>CD4<sup>+</sup> MLN T cells following *H. polygyrus* infection than did wild-type C57BL/6 mice (Fig. 4I), and expression of CD103 among these cells was also higher than in wild-type mice (Fig. 4J). TLR4-deficient mice displayed no differences from wild-type C57BL/6 mice in Foxp3<sup>+</sup>CD4<sup>+</sup> MLN T cell proportions or CD103 expression following *H. polygyrus* infection (Fig. 4E, 4F). MLN total cell counts following *H. polygyrus* infection were comparable to wild-type C57BL/6 mice cell numbers for all genotypes examined (data not shown).

Thus, signaling through individual TLRs contributes differentially to Treg proportions and CD103 expression levels following infection, resulting in no overall differences in Treg phenotype



**FIGURE 2.** Mice singly deficient in specific TLRs harbor *H. polygyrus* burdens similar to those in wild-type C57BL/6 mice. Mice of each genotype were infected with 200 *H. polygyrus* third stage larvae. (A–D) Adult *H. polygyrus* numbers recovered from the intestinal tract 28 d post infection are shown in C57BL/6 and (A) TLR2<sup>−/−</sup> mice, data shown pooled from four independent experiments, each with three to four mice per group; (B) TLR4<sup>−/−</sup> mice, data shown pooled from three independent experiments, each with five to six mice per group; (C) TLR5<sup>−/−</sup> mice, data shown pooled from two independent experiments, each with four to six mice per group; and (D) TLR9<sup>−/−</sup> mice, data shown pooled from two independent experiments, each with five to six mice per group and representative of results from five independent experiments. (E) *H. polygyrus* eggs per gram of feces taken 14 and 28 d post infection in C57BL/6 and TLR2<sup>−/−</sup> mice; data shown at day 14 were pooled from two independent experiments, each with three to four mice per group, and data shown at day 28 were pooled from four independent experiments, each with two to four mice per group. Statistics shown indicate comparisons made between genotypes at the same time point. (F) *H. polygyrus* eggs per gram of feces taken 28 d post infection in C57BL/6, TLR4<sup>−/−</sup>, TLR5<sup>−/−</sup>, and TLR9<sup>−/−</sup> mice; data shown pooled from two to three independent experiments, each with four to six mice per group. \**p* ≤ 0.05.



from wild-type C57BL/6 mice when TLR signaling is ablated in MyD88-deficient mice.

#### TRIF modulates granuloma formation but not antiparasite immunity

MyD88<sup>−/−</sup> mice retain the ability to respond to certain bacterial signals, as TLR4, which recognizes LPS (32), can signal independently of MyD88 through the TRIF adapter protein (33). Thus, only mice deficient in both MyD88 and TRIF are unable to respond to bacterial ligands through TLRs; hence we tested TRIF-deficient and MyD88 × TRIF doubly deficient mice for their susceptibility to *H. polygyrus* infection. Unlike MyD88<sup>−/−</sup> mice, TRIF<sup>−/−</sup> mice did not show increased parasite expulsion, with adult worm numbers (Fig. 5A) and egg burdens (Fig. 5B) matching those of the C57BL/6 wild-type. In contrast, MyD88<sup>−/−</sup>TRIF<sup>−/−</sup> mice were similar to their MyD88<sup>−/−</sup> counterparts, with reduced *H. polygyrus* adult worm survival (Fig. 5C) and egg production (Fig. 5D), compared with wild-type mice, by day 28 post infection, confirming that MyD88 is a key adapter protein in host susceptibility to *H. polygyrus*.

In these experiments, individual TLR-deficient and TRIF-deficient mice were also examined for the presence of granulomas following *H. polygyrus* infection. No mice deficient in individual TLR or TRIF molecules produced the high number of granulomas seen in MyD88<sup>−/−</sup> mice (Fig. 5E). Surprisingly, MyD88<sup>−/−</sup>TRIF<sup>−/−</sup> mice developed far fewer granulomas than mice lacking only MyD88 (Fig. 5E), suggesting that TRIF is required for the high numbers of granulomas observed in the absence of MyD88 or that it interacts with the pathways affected by loss of MyD88.

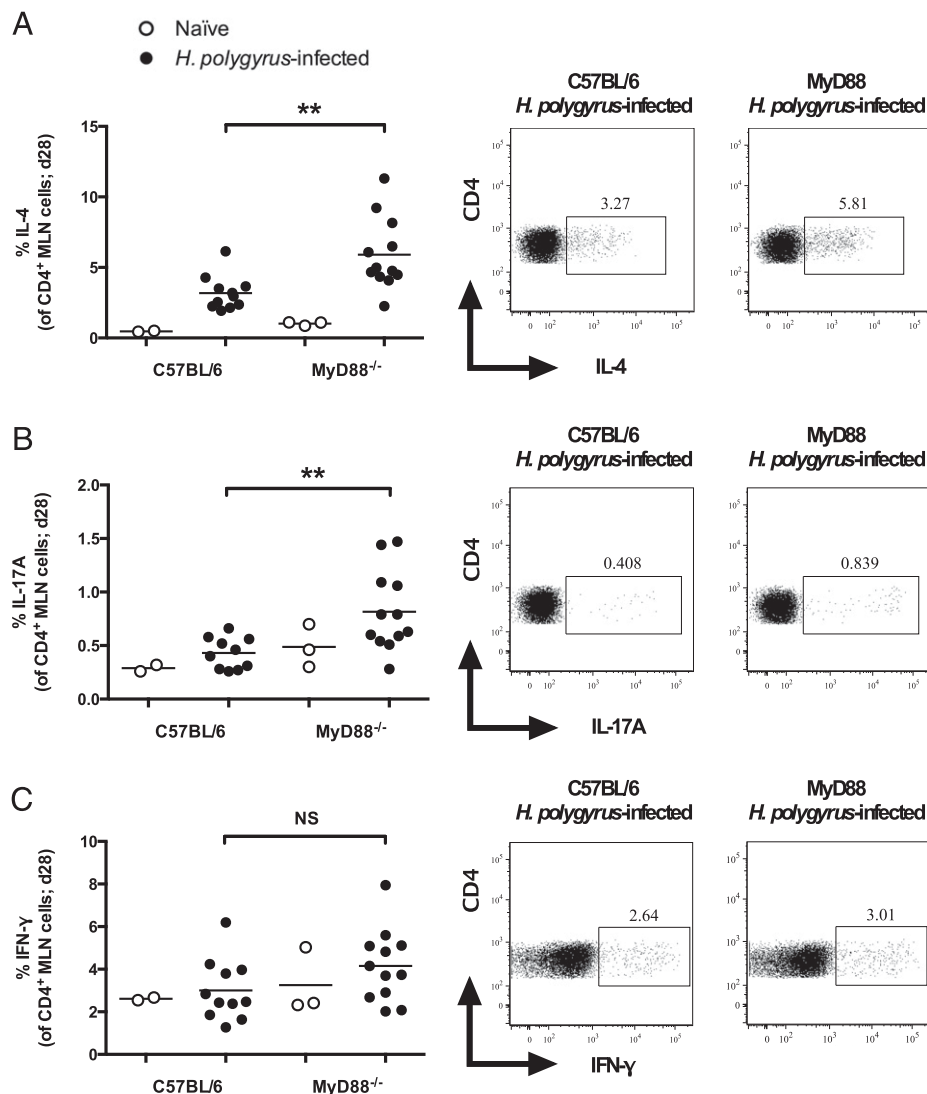
#### Heightened granuloma numbers in MyD88-deficient mice, but not heightened expulsion, can be attributed to a lack of IL-1R signaling

In addition to mediating signaling from TLRs, MyD88 is also a critical adapter protein for signaling by the IL-1 family members IL-1α, IL-1β, IL-18, and IL-33 (28). Because individual TLR-deficient mice did not recapitulate the increased resistance of MyD88-deficient mice to *H. polygyrus*, and IL-1 has recently been reported to promote susceptibility to this parasite (34), we next examined whether mice lacking IL-1R signaling (IL-1R1<sup>−/−</sup>) showed heightened immunity to this parasite. Of interest, IL-1R1<sup>−/−</sup> mice, consistent with previously reported findings in IL-1β<sup>−/−</sup> mice (34), produced high numbers of granulomas in response to infection (Fig. 6A). However, adult *H. polygyrus* burdens 28 d post infection, and egg output at days 14 and 28 post infection, were similar between IL-1R1<sup>−/−</sup> and wild-type C57BL/6 mice (Fig. 6B–D), suggesting that heightened granuloma formation does not contribute to increased parasite expulsion and that the phenotype of IL-1R1 deficiency is not as profound as that reported for mice lacking IL-1β (34).

#### Type 1 IFN signaling may act as downstream inhibitor of granuloma formation

Type 1 IFNs (IFN-α/IFN-β) can be produced in response to stimulation of both MyD88- and TRIF-dependent pathways (35), and signal through a heterodimeric receptor consisting of IFN-αR1 (i.e., IFNAR1) and IFN-αR2c (36). To test whether granuloma formation was affected by a loss of type 1 IFN signaling, we infected IFNAR1<sup>−/−</sup> mice (37) with *H. polygyrus*. IFNAR1<sup>−/−</sup> mice did not share the heightened parasite immunity phenotype shown by MyD88<sup>−/−</sup> mice, as similar numbers of adult worms

**FIGURE 3.** MyD88<sup>-/-</sup> mice mount stronger CD4<sup>+</sup> T cell IL-4 and IL-17A responses following *H. polygyrus* infection. C57BL/6 and MyD88<sup>-/-</sup> mice were left naive or infected with 200 *H. polygyrus* third stage larvae. At 28 d following infection, MLN cells were isolated and restimulated with PMA/ionomycin and brefeldin A, after which cells were stained as indicated and cytokine production was measured by flow cytometry. **(A)** Percentage of IL-4-producing cells among CD4<sup>+</sup>, live lymphocyte cells, and representative dot plots. **(B)** Percentage of IL-17A-producing cells among CD4<sup>+</sup>, live lymphocyte cells, and representative dot plots. **(C)** Percentage of IFN- $\gamma$ -producing cells among CD4<sup>+</sup>, live lymphocyte cells, and representative dot plots. Data shown in (A)–(C) are pooled from two independent experiments, each with four to seven *H. polygyrus*-infected mice per group; naive mice were examined in only one of these experiments. Statistics shown indicate comparisons between infected groups. \*\* $p \leq 0.01$ .



were recovered 28 d post infection as in wild-type C57BL/6 mice (Fig. 7A), and *H. polygyrus* egg production was even greater in IFNAR1<sup>-/-</sup> mice than in wild-type controls 28 d following infection (Fig. 7B). However, this genotype showed increased formation of granulomas (Fig. 7C), indicating that like MyD88, type 1 IFNs inhibit this process in wild-type mice.

## Discussion

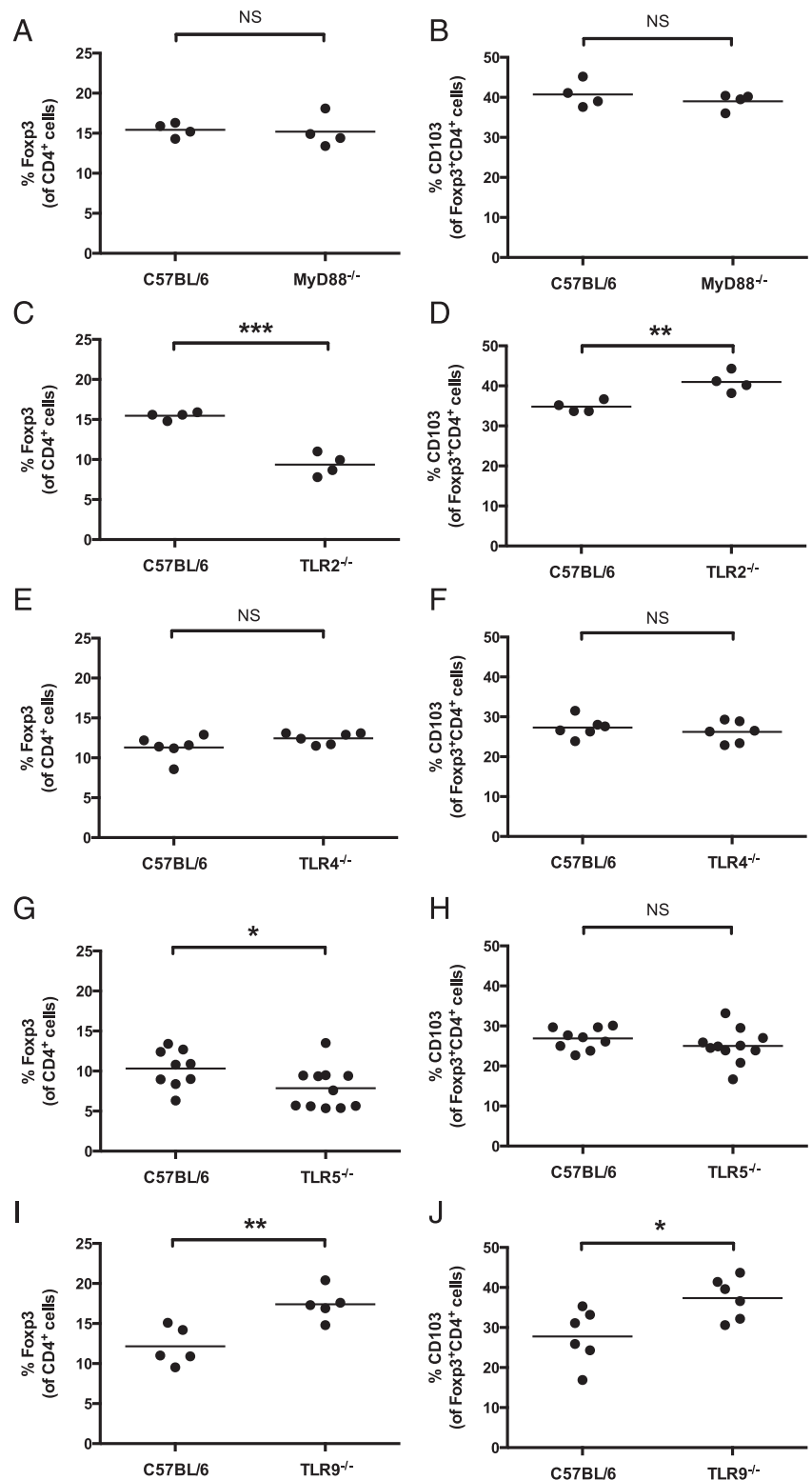
In this study, we investigated the outcome of a primary *H. polygyrus* infection in mice deficient in MyD88 signaling, and found that they were more resistant to infection than wild-type C57BL/6 mice and had a higher frequency of IL-4-producing CD4<sup>+</sup> T cells following infection. In addition, MyD88-deficient mice developed large numbers of granulomas along the intestinal wall in response to infection. Intestinal granulomas that form in response to *H. polygyrus* infection are enriched in alternatively activated macrophages and neutrophils (10, 16), and are more frequent in more resistant strains of mice (10, 19).

Many helminth parasites are able to establish chronic infections in mammals by manipulation of host immunity, which is largely attributed to the secretion and action of parasite excretory-secretory proteins (38). One pathway by which helminths can modulate host signaling is through interference with mammalian TLR signaling (39–41). Several examples of helminth products that can bind to TLRs and mediate signaling through these

receptors have been documented, such as dsRNA from *Schistosoma mansoni* eggs, which binds TLR3, leading to NF- $\kappa$ B activation (42), and the lipid lysophosphatidylserine extracted from *S. mansoni* eggs or adult worms, which stimulates TNF- $\alpha$  and IL-10 production via TLR2 (43). Similarly, the excretory-secretory product ES-62 from the rodent filarial nematode *Acanthocheiloneema viteae* signals via TLR4 to cause IL-12 and TNF- $\alpha$  production (44), and the *S. mansoni* Lewis<sup>X</sup>-containing egg carbohydrate lacto-*N*-fucopentaose III binds TLR4 to drive MAPK activation and IL-4 production (45). LPS signaling through TLR4 does not typically induce IL-4 production, and how helminth products induce differential cytokine production following TLR ligation/activation is yet to be resolved. A potential mechanism by which helminth products can skew downstream responses of TLR signaling is through the additional engagement of co-receptors (46).

Infection with *H. polygyrus* may result in TLR signaling not only through secretory molecules but also by increasing the exposure of immune cells to bacterial ligands. In particular, when *H. polygyrus* larvae disrupt the epithelial cell barrier during migration through to the submucosa, they likely facilitate bacterial translocation and contact with host cells through to the serosal layer of the gut. If so, signals from bacterial ligands may significantly impair the ability of the host to mount a protective response to helminth infection.

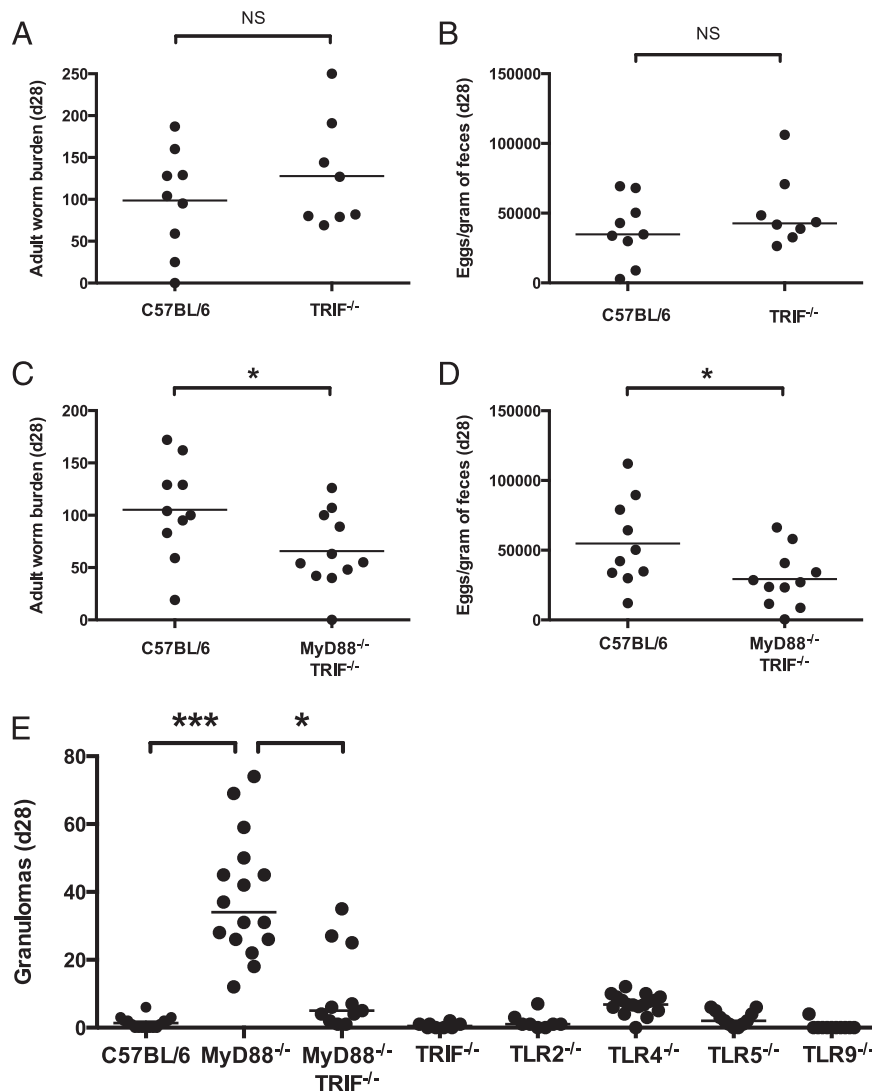
**FIGURE 4.** TLR signaling contributes differentially to MLN Treg proportions following *H. polygyrus* infection. All mice were infected with 200 *H. polygyrus* third stage larvae. At 28 d following infection, MLN cells were isolated, stained as indicated, and analyzed by flow cytometry. **(A, C, E, G, I)** Percent of Foxp3<sup>+</sup> cells among CD4<sup>+</sup> lymphocytes in wild-type C57BL/6 and **(A)** MyD88<sup>-/-</sup>, **(C)** TLR2<sup>-/-</sup>, **(E)** TLR4<sup>-/-</sup>, **(G)** TLR5<sup>-/-</sup>, and **(I)** TLR9<sup>-/-</sup> mice. **(B, D, F, H, J)** Percent of CD103<sup>+</sup> cells among Foxp3<sup>+</sup>CD4<sup>+</sup> lymphocytes in wild-type C57BL/6 and **(B)** MyD88<sup>-/-</sup>, **(D)** TLR2<sup>-/-</sup>, **(F)** TLR4<sup>-/-</sup>, **(H)** TLR5<sup>-/-</sup>, and **(J)** TLR9<sup>-/-</sup> mice. Data shown in **(A)** and **(B)** are representative of results from three independent experiments, each with four to seven mice per group; data in **(C)** and **(D)** are representative of results from two independent experiments, each with four mice per group; data in **(E)** and **(F)** from one experiment with six mice per group; data in **(G)** and **(H)** are pooled from two independent experiments, each with four to six mice per group; and data in **(I)** and **(J)** are representative of results from five independent experiments, each with five to seven mice per group. \**p* ≤ 0.05, \*\**p* ≤ 0.01, \*\*\**p* ≤ 0.001.



The increased *H. polygyrus* expulsion seen in MyD88-deficient mice was not replicated in any individual TLR-deficient mouse examined, although *H. polygyrus* egg output was reduced by day 28 post infection in TLR2-deficient mice, suggesting that signaling through this receptor partially regulates immunity to *H. polygyrus*. Deficiencies in individual TLRs did, however, alter Treg proportions following infection. Consistent with the results described in this article, TLR2 and TLR5 engagement has been previously shown to expand Tregs (47–49), whereas TLR9 stim-

ulation has been shown to limit Treg function (50). TLR4 stimulation enhances Treg survival and proliferation in vitro (51), but in these in vivo experiments, TLR4 deficiency did not affect Treg proportions. Infection status likely determines the importance of TLR signaling in controlling Treg cell frequencies; TLR2-deficient mice have equal proportions of Foxp3<sup>+</sup>CD4<sup>+</sup> cells at the MLN site at steady state (52), yet during the *H. polygyrus* infections performed in this study, and during *Candida albicans* infection (53), proportions are reduced, compared with

**FIGURE 5.** MyD88<sup>-/-</sup>, but not MyD88<sup>-/-</sup> TRIF<sup>-/-</sup>, mice display heightened granuloma formation in response to *H. polygyrus* infection. Mice of each genotype were infected with 200 *H. polygyrus* third stage larvae. (A) Adult *H. polygyrus* numbers recovered from the intestinal tract of C57BL/6 and TRIF<sup>-/-</sup> mice 28 d post infection. (B) *H. polygyrus* eggs per gram of feces from C57BL/6 and TRIF<sup>-/-</sup> mice taken 28 d post infection. (A and B) Data shown pooled from two independent experiments, each with three to five mice per group. (C) Adult *H. polygyrus* numbers recovered from the intestinal tract of C57BL/6 and MyD88<sup>-/-</sup>TRIF<sup>-/-</sup> mice 28 d post infection. (D) *H. polygyrus* eggs per gram of feces from C57BL/6 and MyD88<sup>-/-</sup>TRIF<sup>-/-</sup> mice taken at 28 d post infection. (C and D) Data shown pooled from three independent experiments, each with two to six mice per group. (E) Number of granulomas formed along the small intestinal tract in the indicated mouse genotypes 28 d following *H. polygyrus* infection. Data shown for each genotype are pooled from two or three independent experiments, and experiments for each genotype were performed separately. A statistical comparison is shown only between the C57BL/6, MyD88<sup>-/-</sup>, and MyD88<sup>-/-</sup>TRIF<sup>-/-</sup> groups. \* $p \leq 0.05$ , \*\*\* $p \leq 0.001$ .



wild-type mice. These findings highlight that TLR control of Treg expansion and proliferation is dependent on both site and context (54).

Although no individual TLR gene deficiency recapitulated the phenotype of the MyD88<sup>-/-</sup> mouse, we found that mice deficient in IL-1R1, which signals through MyD88, did reproduce the granulomatous response following *H. polygyrus* infection, suggesting that signaling through the IL-1R1 represses granuloma formation in MyD88-sufficient mice. Consistent with this finding, IL-1 $\beta$ -deficient mice have been reported to develop more, and longer-lasting, granulomas during *H. polygyrus* infection (34). However, despite their enhanced granulomas, IL-1R1<sup>-/-</sup> mice displayed no differences from wild-type mice in adult worm expulsion, indicating that granuloma formation is not a key element of immunity to this parasite. The possibility cannot be excluded, however, that the cellular components of *H. polygyrus*-induced granulomas differ in quality between MyD88-deficient and IL-1R1-deficient mice.

If granulomas are not a mechanism of immunity to *H. polygyrus*, they may instead form to repair damage caused by either the migrating parasite or the resultant immune response (55). Together, these findings lead to the intriguing hypothesis that a component of the microbiota, signaling via IL-1R and MyD88, may function to inhibit granuloma formation in response to helminth infection. The composition of the intestinal microbiota may

thus be a factor in controlling a granulomatous response to *H. polygyrus* infection.

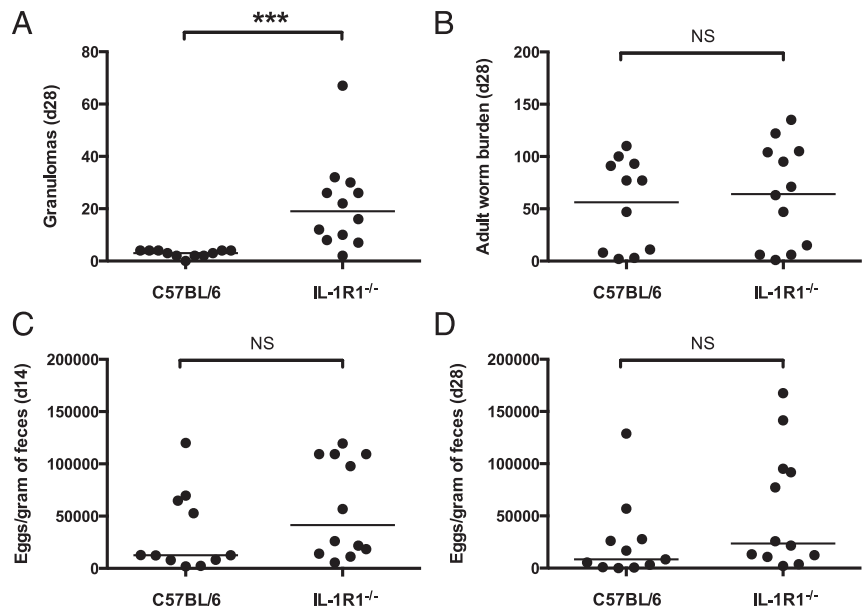
These data also raise broader questions about the role of IL-1 signaling during helminth infections, as IL-1 $\alpha$  and IL-1 $\beta$  appear to promote protective Th2 responses against the helminth parasite *Trichuris muris* (56), yet the IL-1R is reportedly not necessary for protective immunity (57). Moreover, a recent study reported that IL-1 $\beta$ -deficient mice are more resistant to *H. polygyrus* (34), again indicating that a ligand-dependent, but IL-1R-independent, pathway yet to be defined is important in immunity to helminths.

The IL-1 family members IL-18 and IL-33 also signal in a manner that is dependent on MyD88 (28, 58). IL-33 has previously been shown to promote type 2 cytokine production and helminth parasite expulsion (59–61); therefore, it is unlikely that a lack of IL-33 is responsible for the heightened immunity of MyD88<sup>-/-</sup> mice. In fact, IL-33R (ST2)-deficient mice show *H. polygyrus* worm burdens similar to those in wild-type BALB/c controls (34), confirming that this alarmin cytokine is not essential for immunity to *H. polygyrus*, although important in maximizing rejection of other intestinal helminths.

It will be important, however, for future work to examine whether a lack of IL-18 recapitulates the heightened resistance of MyD88<sup>-/-</sup> mice, as during infection with *T. muris*, IL-18 has been shown to promote susceptibility to infection through the



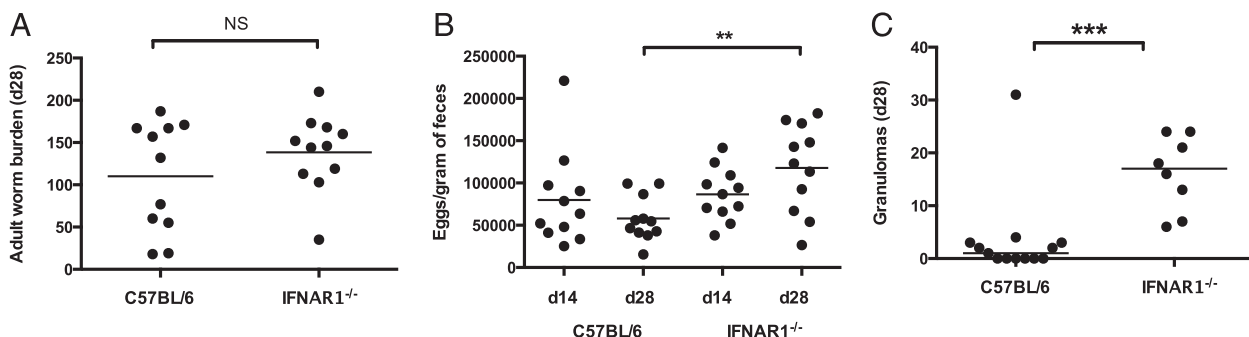
**FIGURE 6.** Heightened granuloma numbers, but not heightened expulsion, can be attributed to a lack of IL-1R1 signaling. C57BL/6 and IL-1R1<sup>-/-</sup> mice were infected with 200 *H. polygyrus* third stage larvae. Data shown are pooled from two independent experiments, each with five to six mice per group. (A) Number of granulomas formed along the small intestinal tract 28 d following *H. polygyrus* infection. (B) Adult *H. polygyrus* numbers recovered from the intestinal tract 28 d post infection. (C) *H. polygyrus* eggs per gram of feces taken 14 d post infection. (D) *H. polygyrus* eggs per gram of feces taken 28 d post infection. \*\*\* $p \leq 0.001$ .



inhibition of Th2 cytokines (62). In some contexts, however, IL-18 has been shown to promote Th2 cytokine production (59), highlighting that the actions of this cytokine are highly dependent on the surrounding cytokine environment and on the genetic background of the host (63, 64). Signaling through inflammasome complexes is required for the release of mature IL-1 and IL-18 (65); thus, modulation of inflammasome signaling to control IL-1 or IL-18 release may be a mechanism by which the microbiota could influence the immune response to helminth infections.

It has previously been reported that DCs unable to signal using MyD88 are impaired in the production of type 1 IFNs (66). We show that mice deficient in type 1 IFN signaling (IFNAR1<sup>-/-</sup> mice) produce the high levels of intestinal granulomas also seen in MyD88<sup>-/-</sup> mice, raising the possibility that following MyD88 stimulation, type 1 IFNs signal via IFNAR1 to inhibit granuloma formation in wild-type mice. Intriguingly, we also show that ablation of signaling through the adapter protein TRIF, on a MyD88<sup>-/-</sup> background, abolished the heightened granuloma formation, suggesting that stimulation of TRIF promotes the formation of intestinal granulomas, through an as-yet-uncharacterized pathway.

With all this information taken together, this report demonstrates the importance of both TLR-dependent and -independent signals through MyD88 in the control of host resistance following helminth infection. A previous report demonstrated that mice deficient in MyD88 show heightened expulsion of *T. muris* (67), suggesting that inhibition of immunity by MyD88-mediated signals may be a common mechanism to allow intestinal colonization by helminth parasites. An absence of MyD88 signaling in our study resulted in heightened *H. polygyrus* expulsion, a phenotype that was partially replicated in TLR2-deficient mice, but not in mice deficient in TLR4, TLR5, or TLR9. This observation raises two possibilities: that signals through individual TLRs redundantly maintain helminth susceptibility or that a TLR-independent pathway is key to the heightened immunity of MyD88-deficient mice. Owing to recent evidence that the presence of the intestinal microbiota is key to maintaining the lifecycle and establishment of helminth parasites within mammalian hosts (68), and with new data showing *H. polygyrus* infection is enhanced by specific commensal bacteria (69), it will be critical to further understand the signaling pathways by which interactions between the microbiota, the immune system, and parasites are controlled.



**FIGURE 7.** Type 1 IFN signaling represses granuloma formation. All mice were infected with 200 *H. polygyrus* third stage larvae. (A) Adult *H. polygyrus* numbers recovered from the intestinal tract of C57BL/6 and IFNAR1<sup>-/-</sup> mice 28 d post infection; data shown are representative of two experiments, each with five to six mice per group. (B) *H. polygyrus* eggs per gram of feces taken 14 and 28 d post infection in C57BL/6 and IFNAR1<sup>-/-</sup> mice; data shown are representative of two experiments, each with five to six mice per group. (C) Number of granulomas formed along the small intestinal tract 28 d following *H. polygyrus* infection. Data shown are pooled from two experiments, each with two to six mice per group. \*\* $p \leq 0.01$ , \*\*\* $p \leq 0.001$ .

## Acknowledgments

We thank Kevin Maloy, Sir William Dunn School of Pathology, University of Oxford, for generous assistance with gene-deficient mice.

## Disclosures

The authors have no financial conflicts of interest.

## References

- Hotez, P. J., P. J. Brindley, J. M. Bethony, C. H. King, E. J. Pearce, and J. Jacobson. 2008. Helminth infections: the great neglected tropical diseases. *J. Clin. Invest.* 118: 1311–1321.
- Behnke, J. M., D. M. Menge, and H. Noyes. 2009. *Heligmosomoides bakeri*: a model for exploring the biology and genetics of resistance to chronic gastrointestinal nematode infections. *Parasitology* 136: 1565–1580.
- Patel, N., T. Kreider, J. F. Urban, Jr., and W. C. Gause. 2009. Characterisation of effector mechanisms at the host:parasite interface during the immune response to tissue-dwelling intestinal nematode parasites. *Int. J. Parasitol.* 39: 13–21.
- Reynolds, L. A., K. J. Filbey, and R. M. Maizels. 2012. Immunity to the model intestinal helminth parasite *Heligmosomoides polygyrus*. *Semin. Immunopathol.* 34: 829–846.
- Grencis, R. K., N. E. Humphreys, and A. J. Bancroft. 2014. Immunity to gastrointestinal nematodes: mechanisms and myths. *Immunol. Rev.* 260: 183–205.
- Wilson, M. S., M. D. Taylor, A. Balic, C. A. M. Finney, J. R. Lamb, and R. M. Maizels. 2005. Suppression of allergic airway inflammation by helminth-induced regulatory T cells. *J. Exp. Med.* 202: 1199–1212.
- Finney, C. A. M., M. D. Taylor, M. S. Wilson, and R. M. Maizels. 2007. Expansion and activation of CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells in *Heligmosomoides polygyrus* infection. *Eur. J. Immunol.* 37: 1874–1886.
- Rausch, S., J. Huehn, C. Loddenkemper, M. R. Hepworth, C. Klotz, T. Sparwasser, A. Hamann, R. Lucius, and S. Hartmann. 2009. Establishment of nematode infection despite increased Th2 responses and immunopathology after selective depletion of Foxp3<sup>+</sup> cells. *Eur. J. Immunol.* 39: 3066–3077.
- Grainger, J. R., K. A. Smith, J. P. Hewitson, H. J. McSorley, Y. Hargus, K. J. Filbey, C. A. Finney, E. J. Greenwood, D. P. Knox, M. S. Wilson, et al. 2010. Helminth secretions induce de novo T cell Foxp3 expression and regulatory function through the TGF- $\beta$  pathway. *J. Exp. Med.* 207: 2331–2341.
- Filbey, K. J., J. R. Grainger, K. A. Smith, L. Boon, N. van Rooijen, Y. Hargus, S. Jenkins, J. P. Hewitson, and R. M. Maizels. 2014. Innate and adaptive type 2 immune cell responses in genetically controlled resistance to intestinal helminth infection. *Immunol. Cell Biol.* 92: 436–448.
- Mitchell, G. F., and S. J. Prowse. 1979. Three consequences of infection with *Nematostrioides dubius* in three inbred strains of mice. *J. Parasitol.* 65: 820–822.
- Wahid, F. N., M. Robinson, and J. M. Behnke. 1989. Immunological relationships during primary infection with *Heligmosomoides polygyrus* (*Nematostrioides dubius*): expulsion of adult worms from fast responder syngeneic and hybrid strains of mice. *Parasitology* 98: 459–469.
- Hewitson, J. P., K. J. Filbey, J. R. Grainger, A. A. Dowle, M. Pearson, J. Murray, Y. Hargus, and R. M. Maizels. 2011. *Heligmosomoides polygyrus* elicits a dominant nonprotective antibody response directed against restricted glycan and peptide epitopes. *J. Immunol.* 187: 4764–4777.
- Hewitson, J. P., A. C. Ivens, Y. Hargus, K. J. Filbey, H. J. McSorley, J. Murray, S. Bridgett, D. Ashford, A. A. Dowle, and R. M. Maizels. 2013. Secretion of protective antigens by tissue-stage nematode larvae revealed by proteomic analysis and vaccination-induced sterile immunity. *PLoS Pathog.* 9: e1003492.
- Behnke, J. M., and M. Robinson. 1985. Genetic control of immunity to *Nematostrioides dubius*: a 9-day anthelmintic abbreviated immunizing regime which separates weak and strong responder strains of mice. *Parasite Immunol.* 7: 235–253.
- Anthony, R. M., J. F. Urban, Jr., F. Alem, H. A. Hamed, C. T. Roza, J. L. Boucher, N. Van Rooijen, and W. C. Gause. 2006. Memory T<sub>H</sub>2 cells induce alternatively activated macrophages to mediate protection against nematode parasites. *Nat. Med.* 12: 955–960.
- Cywińska, A., K. Czumińska, and A. Schollenberger. 2004. Granulomatous inflammation during *Heligmosomoides polygyrus* primary infections in FVB mice. *J. Helminthol.* 78: 17–24.
- Morimoto, M., M. Morimoto, J. Whitmire, S. Xiao, R. M. Anthony, H. Mirakami, R. A. Star, J. F. Urban, Jr., and W. C. Gause. 2004. Peripheral CD4 T cells rapidly accumulate at the host:parasite interface during an inflammatory Th2 memory response. *J. Immunol.* 172: 2424–2430.
- Menge, D. M., J. M. Behnke, A. Lowe, J. P. Gibson, F. A. Iraqi, R. L. Baker, and D. Wakelin. 2003. Mapping of chromosomal regions influencing immunological responses to gastrointestinal nematode infections in mice. *Parasite Immunol.* 25: 341–349.
- Wescott, R. B. 1968. Experimental *Nematostrioides dubius* infection in germ-free and conventional mice. *Exp. Parasitol.* 22: 245–249.
- Weinstein, P. P., W. L. Newton, T. K. Sawyer, and R. I. Sommerville. 1969. *Nematostrioides dubius*: development and passage in the germfree mouse, and a comparative study of the free-living stages in germfree feces and conventional cultures. *Trans. Am. Microsc. Soc.* 88: 95–117.
- Chang, J., and R. B. Wescott. 1972. Infectivity, fecundity, and survival of *Nematostrioides dubius* in gnotobiotic mice. *Exp. Parasitol.* 32: 327–334.
- Rakoff-Nahoum, S., J. Paglino, F. Eslami-Varzaneh, S. Edberg, and R. Medzhitov. 2004. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell* 118: 229–241.
- Elinav, E., T. Strowig, A. L. Kau, J. Henao-Mejia, C. A. Thaiss, C. J. Booth, D. R. Peaper, J. Bertin, S. C. Eisenbarth, J. I. Gordon, and R. A. Flavell. 2011. NLRP6 inflammasome regulates colonic microbial ecology and risk for colitis. *Cell* 145: 745–757.
- Kirkland, D., A. Benson, J. Mirpuri, R. Pifer, B. Hou, A. L. DeFranco, and F. Yarovinsky. 2012. B cell-intrinsic MyD88 signaling prevents the lethal dissemination of commensal bacteria during colonic damage. *Immunity* 36: 228–238.
- Hand, T. W., L. M. Dos Santos, N. Bouladoux, M. J. Molloy, A. J. Pagán, M. Pepper, C. L. Maynard, C. O. Elson, III, and Y. Belkaid. 2012. Acute gastrointestinal infection induces long-lived microbiota-specific T cell responses. *Science* 337: 1553–1556.
- Medzhitov, R., P. Preston-Hurlburt, E. Kopp, A. Stadler, C. Chen, S. Ghosh, and C. A. Janeway, Jr. 1998. MyD88 is an adaptor protein in the hToll/IL-1 receptor family signaling pathways. *Mol. Cell* 2: 253–258.
- Adachi, O., T. Kawai, K. Takeda, M. Matsumoto, H. Tsutsui, M. Sakagami, K. Nakanishi, and S. Akira. 1998. Targeted disruption of the MyD88 gene results in loss of IL-1- and IL-18-mediated function. *Immunity* 9: 143–150.
- Urban, J. F., Jr., K. B. Madden, A. Svetić, A. Cheever, P. P. Trotta, W. C. Gause, I. M. Katona, and F. D. Finkelman. 1992. The importance of Th2 cytokines in protective immunity to nematodes. *Immunol. Rev.* 127: 205–220.
- Urban, J. F., Jr., I. M. Katona, W. E. Paul, and F. D. Finkelman. 1991. Interleukin 4 is important in protective immunity to a gastrointestinal nematode infection in mice. *Proc. Natl. Acad. Sci. USA* 88: 5513–5517.
- Smith, K. A., and R. M. Maizels. 2014. IL-6 controls susceptibility to helminth infection by impeding Th2 responsiveness and altering the Treg phenotype in vivo. *Eur. J. Immunol.* 44: 150–161.
- Poltorak, A., X. He, I. Smirnova, M. Y. Liu, C. Van Huffel, X. Du, D. Birdwell, E. Alejos, M. Silva, C. Galanos, et al. 1998. Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in Tlr4 gene. *Science* 282: 2085–2088.
- Yamamoto, M., S. Sato, H. Hemmi, K. Hoshino, T. Kaisho, H. Sanjo, O. Takeuchi, M. Sugiyama, M. Okabe, K. Takeda, and S. Akira. 2003. Role of adaptor TRIF in the MyD88-independent toll-like receptor signaling pathway. *Science* 301: 640–643.
- Zaiss, M. M., K. M. Maslowski, I. Mosconi, N. Guenat, B. J. Marsland, and N. L. Harris. 2013. IL-1 $\beta$  suppresses innate IL-25 and IL-33 production and maintains helminth chronicity. *PLoS Pathog.* 9: e1003531.
- Trinchieri, G. 2010. Type I interferon: friend or foe? *J. Exp. Med.* 207: 2053–2063.
- Pestka, S., C. D. Krause, and M. R. Walter. 2004. Interferons, interferon-like cytokines, and their receptors. *Immunol. Rev.* 202: 8–32.
- Hwang, S. Y., P. J. Hertzog, K. A. Holland, S. H. Sumarsono, M. J. Tymms, J. A. Hamilton, G. Whitty, I. Bertonecello, and I. Kola. 1995. A null mutation in the gene encoding a type I interferon receptor component eliminates anti-proliferative and antiviral responses to interferons  $\alpha$  and  $\beta$  and alters macrophage responses. *Proc. Natl. Acad. Sci. USA* 92: 11284–11288.
- Hewitson, J. P., J. R. Grainger, and R. M. Maizels. 2009. Helminth immunoregulation: the role of parasite secreted proteins in modulating host immunity. *Mol. Biochem. Parasitol.* 167: 1–11.
- Semnani, R. T., P. G. Venugopal, C. A. Leifer, S. Mostböck, H. Sabzevari, and T. B. Nutman. 2008. Inhibition of TLR3 and TLR4 function and expression in human dendritic cells by helminth parasites. *Blood* 112: 1290–1298.
- Ludwig-Portugall, I., and L. E. Layland. 2012. TLRs, Treg, and B cells, an interplay of regulation during helminth infection. *Front. Immunol.* 3: 8.
- Kane, C. M., L. Cervi, J. Sun, A. S. McKee, K. S. Masek, S. Shapira, C. A. Hunter, and E. J. Pearce. 2004. Helminth antigens modulate TLR-initiated dendritic cell activation. *J. Immunol.* 173: 7454–7461.
- Aksoy, E., C. S. Zouain, F. Vanhoutte, J. Fontaine, N. Pavelka, N. Thieblemont, F. Willems, P. Ricciardi-Castagnoli, M. Goldman, M. Capron, et al. 2005. Double-stranded RNAs from the helminth parasite *Schistosoma* activate TLR3 in dendritic cells. *J. Biol. Chem.* 280: 277–283.
- van der Kleij, D., E. Latz, J. F. Brouwers, Y. C. Kruijs, M. Schmitz, E. A. Kurt-Jones, T. Espevik, E. C. de Jong, M. L. Kapsenberg, D. T. Golenbock, et al. 2002. A novel host-parasite lipid cross-talk. Schistosomal lyso-phosphatidylserine activates toll-like receptor 2 and affects immune polarization. *J. Biol. Chem.* 277: 48122–48129.
- Goodridge, H. S., F. A. Marshall, K. J. Else, K. M. Houston, C. Egan, L. Al-Riyami, F. Y. Liew, W. Harnett, and M. M. Harnett. 2005. Immunomodulation via novel use of TLR4 by the filarial nematode phosphorylcholine-containing secreted product, ES-62. *J. Immunol.* 174: 284–293.
- Thomas, P. G., M. R. Carter, O. Atochina, A. A. Da'Dara, D. Piskorska, E. McGuire, and D. A. Harn. 2003. Maturation of dendritic cell 2 phenotype by a helminth glycan uses a Toll-like receptor 4-dependent mechanism. *J. Immunol.* 171: 5837–5841.
- van Riet, E., F. C. Hartgers, and M. Yazdanbakhsh. 2007. Chronic helminth infections induce immunomodulation: consequences and mechanisms. *Immunobiology* 212: 475–490.
- Crellin, N. K., R. V. Garcia, O. Hadisfar, S. E. Allan, T. S. Steiner, and M. K. Levings. 2005. Human CD4<sup>+</sup> T cells express TLR5 and its ligand flagellin enhances the suppressive capacity and expression of FOXP3 in CD4<sup>+</sup>CD25<sup>+</sup> T regulatory cells. *J. Immunol.* 175: 8051–8059.
- Liu, H., M. Komai-Koma, D. Xu, and F. Y. Liew. 2006. Toll-like receptor 2 signaling modulates the functions of CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells. *Proc. Natl. Acad. Sci. USA* 103: 7048–7053.
- Sutmoller, R. P., M. H. den Broek, M. Kramer, E. J. Binnink, L. W. Toonen, B. J. Kullberg, L. A. Joosten, S. Akira, M. G. Netea, and G. J. Adema. 2006. Toll-like receptor 2 controls expansion and function of regulatory T cells. *J. Clin. Invest.* 116: 485–494.

50. Hall, J. A., N. Bouladoux, C. M. Sun, E. A. Wohlfert, R. B. Blank, Q. Zhu, M. E. Grigg, J. A. Berzofsky, and Y. Belkaid. 2008. Commensal DNA limits regulatory T cell conversion and is a natural adjuvant of intestinal immune responses. *Immunity* 29: 637–649.
51. Caramalho, I., T. Lopes-Carvalho, D. Ostler, S. Zelenay, M. Haury, and J. Demengeot. 2003. Regulatory T cells selectively express toll-like receptors and are activated by lipopolysaccharide. *J. Exp. Med.* 197: 403–411.
52. Boulard, O., M. J. Asquith, F. Powrie, and K. J. Maloy. 2010. TLR2-independent induction and regulation of chronic intestinal inflammation. *Eur. J. Immunol.* 40: 516–524.
53. Netea, M. G., R. Sutmoller, C. Hermann, C. A. A. Van der Graaf, J. W. M. Van der Meer, J. H. van Krieken, T. Hartung, G. Adema, and B. J. Kullberg. 2004. Toll-like receptor 2 suppresses immunity against *Candida albicans* through induction of IL-10 and regulatory T cells. *J. Immunol.* 172: 3712–3718.
54. Grainger, J. R., J. A. Hall, N. Bouladoux, G. Oldenhove, and Y. Belkaid. 2010. Microbe-dendritic cell dialog controls regulatory T-cell fate. *Immunol. Rev.* 234: 305–316.
55. Martin, P., and S. J. Leibovich. 2005. Inflammatory cells during wound repair: the good, the bad and the ugly. *Trends Cell Biol.* 15: 599–607.
56. Helmbly, H., and R. K. Grencis. 2004. Interleukin 1 plays a major role in the development of Th2-mediated immunity. *Eur. J. Immunol.* 34: 3674–3681.
57. Humphreys, N. E., and R. K. Grencis. 2009. IL-1-dependent, IL-1R1-independent resistance to gastrointestinal nematodes. *Eur. J. Immunol.* 39: 1036–1045.
58. Brint, E. K., D. Xu, H. Liu, A. Dunne, A. N. McKenzie, L. A. O'Neill, and F. Y. Liew. 2004. ST2 is an inhibitor of interleukin 1 receptor and Toll-like receptor 4 signaling and maintains endotoxin tolerance. *Nat. Immunol.* 5: 373–379.
59. Kondo, Y., T. Yoshimoto, K. Yasuda, S. Futatsugi-Yumikura, M. Morimoto, N. Hayashi, T. Hoshino, J. Fujimoto, and K. Nakanishi. 2008. Administration of IL-33 induces airway hyperresponsiveness and goblet cell hyperplasia in the lungs in the absence of adaptive immune system. *Int. Immunol.* 20: 791–800.
60. Neill, D. R., S. H. Wong, A. Bellosi, R. J. Flynn, M. Daly, T. K. Langford, C. Bucks, C. M. Kane, P. G. Fallon, R. Pannell, et al. 2010. Nuocytes represent a new innate effector leukocyte that mediates type-2 immunity. *Nature* 464: 1367–1370.
61. Humphreys, N. E., D. Xu, M. R. Hepworth, F. Y. Liew, and R. K. Grencis. 2008. IL-33, a potent inducer of adaptive immunity to intestinal nematodes. *J. Immunol.* 180: 2443–2449.
62. Helmbly, H., K. Takeda, S. Akira, and R. K. Grencis. 2001. Interleukin (IL)-18 promotes the development of chronic gastrointestinal helminth infection by downregulating IL-13. *J. Exp. Med.* 194: 355–364.
63. Nakanishi, K., T. Yoshimoto, H. Tsutsui, and H. Okamura. 2001. Interleukin-18 regulates both Th1 and Th2 responses. *Annu. Rev. Immunol.* 19: 423–474.
64. Xu, D., V. Trajkovic, D. Hunter, B. P. Leung, K. Schulz, J. A. Gracie, I. B. McInnes, and F. Y. Liew. 2000. IL-18 induces the differentiation of Th1 or Th2 cells depending upon cytokine milieu and genetic background. *Eur. J. Immunol.* 30: 3147–3156.
65. Franchi, L., R. Muñoz-Planillo, and G. Núñez. 2012. Sensing and reacting to microbes through the inflammasomes. *Nat. Immunol.* 13: 325–332.
66. Takaki, H., M. Takeda, M. Tahara, M. Shingai, H. Oshiumi, M. Matsumoto, and T. Seya. 2013. The MyD88 pathway in plasmacytoid and CD4<sup>+</sup> dendritic cells primarily triggers type I IFN production against measles virus in a mouse infection model. *J. Immunol.* 191: 4740–4747.
67. Helmbly, H., and R. K. Grencis. 2003. Essential role for TLR4 and MyD88 in the development of chronic intestinal nematode infection. *Eur. J. Immunol.* 33: 2974–2979.
68. Hayes, K. S., A. J. Bancroft, M. Goldrick, C. Portsmouth, I. S. Roberts, and R. K. Grencis. 2010. Exploitation of the intestinal microflora by the parasitic nematode *Trichuris muris*. *Science* 328: 1391–1394.
69. Reynolds, L., K. Smith, K. Filbey, Y. Hargus, J. Hewitson, S. Redpath, Y. Valdez, M. Yebra, B. B. Finlay, and R. Maizels. 2014. Commensal-pathogen interactions in the intestinal tract: lactobacilli promote infection with, and are promoted by, helminth parasites. *Gut Microbes* 5:10–9.