Mutations in DCC cause isolated Agenesis of the Corpus Callosum with incomplete penetrance

3

Ashley PL Marsh^{1,2} [†], Delphine Heron^{3,4,5} [†], Timothy J Edwards^{6,7} [†], Angélique Quartier⁸, 4 Charles Galea⁹, Caroline Nava^{3,10}, Agnès Rastetter¹⁰, Marie-Laure Moutard^{11,12,13}, Vicki 5 Anderson¹⁴, Pierre Bitoun¹⁵, Jens Bunt⁶, Anne Faudet³, Catherine Garel¹⁶, Greta Gillies¹, Ilan 6 Gobius⁶, Justine Guegan¹⁷, Solveig Heide^{3,4}, Boris Keren^{3,10}, Fabien Lesne³, Vesna Lukic¹⁸, 7 Simone A Mandelstam^{2,19,20}, George Mcgillivrav²¹, Alissandra McIlrov¹⁴, Aurélie Méneret^{10,22}, 8 Cyril Mignot^{3,4,5}, Laura R Morcom⁶, Sylvie Odent^{23,24}, Annalisa Paolino⁶, Kate Pope¹, Florence 9 Riant²⁵, Gail A Robinson²⁶, Megan Spencer-Smith^{14,27}, Myriam Srour^{28,29}, Sarah EM 10 Stephenson^{1,2}, Rick Tankard^{30,31}, Oriane Trouillard¹⁰, Quentin Welniarz^{10,32}, Amanda Wood^{14,33}, 11 Alexis Brice,^{3,10} Guy Rouleau^{29,34}, Tania Attié-Bitach³⁵, Martin B Delatycki^{1,2,36} Jean-Louis 12 Mandel^{8,37}, David J Amor^{1,2}, Emmanuel Roze^{10,22}, Amélie Piton^{8,37}, Melanie Bahlo^{30,31}, Thierry 13 Billette de Villemeur^{5,11,12,38}, Elliott H Sherr³⁹, Richard J Leventer^{2,40,41}, Linda J Richards^{6,42}§*, 14 Paul J Lockhart^{1,2} §*, Christel Depienne^{3,8,10,37} §* 15

16

¹ Bruce Lefroy Centre for Genetic Health Research, Murdoch Childrens Research Institute, Royal
Children's Hospital, Parkville, Victoria 3052, Australia.

² Department of Paediatrics, University of Melbourne, Parkville, Victoria 3052, Australia.

20 ³ AP-HP, Hôpital de la Pitié-Salpêtrière, Département de Génétique, F-75013, Paris, France.

⁴ Groupe de Recherche Clinique (GRC) "déficience intellectuelle et autisme" UPMC, 75013

22 Paris, France.

⁵Centre de Référence "déficiences intellectuelles de causes rares", 75013 Paris, France.

⁶ The University of Queensland, Queensland Brain Institute, St Lucia, Brisbane, 4072, Australia.

- ⁷ The University of Queensland, School of Medicine, Herston, Brisbane, 4006, Australia.
- ⁸ Département de Médicine translationnelle et Neurogénétique, IGBMC, CNRS UMR
- 27 7104/INSERM U964/Université de Strasbourg, 67400 Illkirch, France.
- ⁹ Drug Delivery, Disposition and Dynamics (D4), Monash Institute of Pharmaceutical Sciences,
- 29 Monash University, Parkville, Victoria, 3052 Australia.
- ¹⁰ INSERM, U 1127, CNRS UMR 7225, Sorbonne Universités, UPMC Univ Paris 06 UMR S
- 31 1127, Institut du Cerveau et de la Moelle épinière, ICM, F-75013, Paris, France
- 32 ¹¹ AP-HP, Hôpital Trousseau, service de neuropédiatrie, 75012 Paris, France.
- 33 ¹² UPMC, GRC ConCer-LD, Sorbonne université, Paris France.
- ¹³Centre de référence "Neurogénétique", 75013 Paris France.
- ¹⁴ Developmental Imaging and Child Neuropsychology Research groups, Murdoch Childrens
- 36 Research Institute, Parkville, Victoria 3052, Australia.
- 37 ¹⁵ Génétique Médicale, CHU Paris Nord, Hôpital Jean Verdier, 93140 Bondy, France.
- ¹⁶ AP-HP, GHUEP, Hôpital Armand-Trousseau, Service de Radiologie, 75012 Paris, France.
- ¹⁷ iCONICS facility, ICM, 75013, Paris, France.
- 40 ¹⁸ Bioinformatics Division, The Walter and Eliza Hall Institute of Medical Research, 1G Royal
- 41 Parade, Parkville, Victoria 3052, Australia.
- 42 ¹⁹ The Florey Institute of Neuroscience and Mental Health, Melbourne, Victoria, Australia.
- 43 ²⁰ Department of Radiology, University of Melbourne, Royal Children's Hospital, Parkville,
- 44 Victoria 3052, Australia.
- ²¹ Victorian Clinical Genetics Services, Murdoch Childrens Research Institute, Parkville, Victoria
 3052, Australia.
- 47 ²² AP-HP, Hôpital de la Pitié-Salpêtrière, Département de Neurologie, F-75013, Paris, France.

- 48 ²³ Service de Génétique Clinique, Centre de référence CLAD-Ouest, CHU Rennes, 35000
- 49 Rennes, France.
- 50 ²⁴ UMR 6290 CNRS, IGDR Institut de Génétique et développement de Rennes, Université de
- 51 Rennes1, Rennes France.
- 52 ²⁵ AP-HP, Groupe Hospitalier Saint Louis, Lariboisière, Fernand Widal, Laboratoire de
- 53 Génétique, 75010 Paris, France.
- ²⁶Neuropsychology Research Unit, School of Psychology, The University of Queensland,
- 55 Brisbane QLD 4072, Australia.
- 56 ²⁷ School of Psychological Sciences and Monash Institute of Cognitive and Clinical
- 57 Neurosciences, Monash University, Clayton Campus, Clayton Victoria 3800, Australia.
- ²⁸ Department of Pediatrics, Montreal Children's Hospital, McGill University, Montréal, Quebec,
- 59 H4A 3J1, Canada.
- ²⁹ Department of Neurology and Neurosurgery, McGill University Health Center, Montreal,
- 61 Quebec, H3A 2B4 Canada.
- ³⁰ Population Health and Immunity Division, The Walter and Eliza Hall Institute of Medical
- 63 Research, Parkville, Victoria, 3052, Australia.
- ⁶⁴ ³¹ Department of Medical Biology, The University of Melbourne, Parkville, Victoria, 3052,
- 65 Australia.
- ³² Institut de Biologie Paris Seine, Neuroscience Paris Seine, Sorbonne Universités, UPMC Univ
- 67 Paris 06, INSERM, CNRS, F-75005, Paris, France
- ³³ School of Life and Health Sciences, Aston University, Birmingham, B4 7ET, United-Kingdom.
- ⁶⁹ ³⁴ Montreal Neurological Institute and Hospital, McGill University, Montréal, Quebec H3A 2B4,
- 70 Canada.

71	³⁵ INSERM U1163, Laboratory of Embryology and Genetics of Congenital Malformations, Paris
72	Descartes University, Sorbonne Paris Cité and Imagine Institute, 75015 Paris, France;
73	Département de Génétique, Hôpital Necker - Enfants Malades, Assistance Publique - Hôpitaux
74	de Paris, 75015 Paris, France.
75	³⁶ Victorian Clinical Genetics Services, Parkville, Victoria, 3052, Australia.
76	³⁷ Laboratoires de génétique, Institut de génétique médicale d'Alsace, Hôpitaux Universitaires de
77	Strasbourg, 67 000 Strasbourg, France.
78	³⁸ INSERM U1141, 75019 Paris, France.
79	³⁹ Department of Neurology, UCSF Benioff Children's Hospital, San Francisco, California, USA.
80	⁴⁰ Neuroscience Research Group, Murdoch Childrens Research Institute, Parkville, Victoria 3052,
81	Australia.
82	⁴¹ Department of Neurology, University of Melbourne, Royal Children's Hospital, Parkville,
83	Victoria 3052, Australia.
84	⁴² The University of Queensland, School of Biomedical Sciences, St Lucia, Brisbane, 4072,
85	Australia.
86	
87	† These authors contributed equally to this work.
88	§ Equal last and senior authors
89	* Corresponding authors. E-mail: depiennc@igbmc.fr (CD), paul.lockhart@mcri.edu.au (PJL),
90	richards@uq.edu.au (LJR)
91	
92	
93	

- 3 -

94 ABSTRACT

95 Brain malformations involving the corpus callosum are common in children with developmental 96 disabilities. We report that *DCC* mutations cause isolated agenesis of the corpus callosum (ACC), 97 without intellectual disability, in four families and five sporadic individuals. *DCC* mutations 98 result in variable dominant phenotypes with reduced penetrance including mirror movements and 99 ACC associated with a favorable developmental prognosis. Possible phenotype modifiers include 100 the type and location of mutation and sex of the individual.

101

102 MAIN TEXT

103 The corpus callosum (CC) is the main cerebral commissure in placental mammals with a key role 104 in communication between the brain hemispheres¹. Formation of the CC is a complex process 105 involving ligands such as those in the Netrin, Ephrin, Semaphorin and Slit families and their receptors². Agenesis of the corpus callosum (ACC) is the complete or partial absence of the CC. 106 107 This frequent brain malformation affects ~1/4,000 newborns and 3-5% of children with intellectual disability (ID)^{3,4} and is a common cause of late pregnancy termination⁵. Mutations in 108 109 many genes cause syndromes with ID and ACC, whereas the genetics of isolated ACC remain poorly understood^{3,6,7}. The Netrin receptor Dcc plays a critical role in CC development in mice 110 by guiding callosal axons at the midline⁸. While mutations in *DCC* have been associated with 111 congenital mirror movements (MM) in humans⁹, they have not been described in individuals with 112 113 ACC.

114

We investigated four multigenerational families with individuals presenting with ACC, MM or both phenotypes segregating as autosomal dominant traits (Fig. 1a). Neuroimaging and clinical studies confirmed that complete or partial ACC was isolated in most cases (Fig. 1b, Fig. S1) and 118 associated with a range of intellectual abilities (normal-borderline impaired); additionally, 119 specific cognitive impairments, including language delay or visuospatial deficits, were 120 documented (Table S1). Diffusion MRI tractography using probabilistic constrained spherical 121 deconvolution identified reduced crossing of descending corticospinal tract projections at the 122 pyramidal decussation in all affected individuals in families 2 and 4 with either ACC and MM 123 (n=5) or MM only (n=2) (Fig. 1c, Fig. S2). The tractography results for other commissural fibers, 124 including the decussation of the superior cerebellar peduncles, anterior commissure, posterior 125 commissure and optic chiasm, were comparable between the affected individuals and controls.

126

127 Linkage analysis and exome sequencing of three affected individuals in family 1 identified two 128 shared candidate variants in the 16 linkage regions (Fig. S3, Table S2) including a truncating 129 mutation in DCC (NM 005215.3:c.925delA, p.(Thr309ProfsTer26)). For family 2, 48 candidate 130 variants were identified in 28 linkage regions (Fig. S4, Table S3), including a missense variant 131 (c.2378T>G, p.(Val793Gly)) in DCC. The previously-reported nonsense mutation (c.823C>T, p.(Arg275Ter)) in DCC segregated with MM in five individuals of family 3^{10} . Further 132 133 investigation revealed two additional female mutation carriers with ACC and a male carrier with 134 MM who had a thin rostrum. Direct screening in family 4 identified a heterozygous DCC 135 missense variant (c.2414G>A, p.(Gly805Glu)). All four DCC mutations were absent from public 136 databases, including 1000 Genomes and ExAC, and segregated with ACC and/or MM in all 137 available individuals tested (Fig. 1a). In addition, we sequenced DCC in 70 unrelated individuals 138 with ACC including 46 with normal cognitive development. Five individuals, all with isolated 139 complete ACC, had at least one heterozygous missense variant altering a conserved amino acid of 140 DCC (Fig. 1d, Fig. S1 and S5, Table 1). Analysis of all available imaging in mutation-positive 141 individuals with complete ACC also showed absence of the hippocampal commissure and cingulate gyri, and dysmorphic lateral ventricles (usually colpocephaly) as would be expected
(detailed in Online methods). Apart from this, no consistent additional brain malformations were
seen.

145

146 We analyzed the phenotype of individuals with DCC mutations reported in the literature and in 147 this study (Table S4) to assess the penetrance of MM and ACC. Of the 88 individuals with DCC 148 mutations identified to date, 50 had MM; among the 39 who had brain imaging, 19 exhibited 149 ACC. Excluding the index individuals from the analysis, the penetrance of MM was estimated to 150 be 42% and the penetrance of ACC to be 26% (Table S5). Overall, males (n=31) exhibited MM 151 more frequently than females (n=19, male:female ratio=1.8, p=0.0027, Fisher's Exact test; Table 152 S5) while, in individuals with truncating DCC variants, ACC was more often present in females 153 (n=7) than males (n=1, male:female ratio=0.2). Sex differences in CC anatomy have been associated with testosterone levels during prenatal brain development¹¹⁻¹³; therefore, we tested 154 155 the effect of androgens on DCC expression. Independent analysis by RNAseq and RT-qPCR 156 demonstrated a significant dose-dependent increase in DCC expression in human neural stem 157 cells treated with 10 nM or 100 nM testosterone (Fig. S6). Since variants introducing a premature 158 stop codon generally result in haploinsufficiency due to nonsense mediated decay of the mutant 159 mRNA, it is possible that ACC may occur when the amount of *DCC* mRNA/protein falls below a 160 threshold level during CC development, which would occur more frequently in females. 161 However, given the incomplete penetrance observed in both sexes, the phenotypic outcome must 162 also be influenced by additional genetic, epigenetic and/or environmental factors. Interestingly, 163 families 1 and 3, in which a majority of females display ACC, are both of North African 164 background, supporting the hypothesis of genetic modifiers.

165

166 Contrary to truncating variants, missense mutant proteins are usually present in the cell and can 167 interfere with the function of the wildtype protein, potentially resulting in differing phenotypes 168 compared to haploinsufficiency for the same protein. Binding of Netrin-1 to DCC results in 169 intracellular homodimerization or heterodimerization with UNC5, another axon guidance 170 receptor, and is critical for both the chemoattractive and chemorepulsive properties of the signaling complexes^{15,16}. The Netrin-1 binding region involves the 4th, 5th and 6th fibronectin 171 type III-like domains of $DCC^{14,15}$, therefore amino acid substitutions in this binding region may 172 173 compromise DCC function. Five of the eight DCC missense variants identified in individuals 174 with ACC are located in the Netrin-1 binding region (Fig. 1d), which represents a considerable 175 enrichment compared to missense variants located in this domain in ExAC (5/74, 6.7% versus $519/\sim60000$, 0.86%; p=5x10⁻⁴ (all rare variants) or 284/~60000, 0.47%; p=3x10⁻⁵ (rare variants) 176 177 predicted to be damaging by SIFT), Fisher's exact test (Table S6)). Given the reduced penetrance 178 and mild phenotype of DCC-related ACC, it is possible that some individuals described in ExAC 179 have pathogenic DCC mutations and undiagnosed ACC.

180

181 Modelling of DCC missense variants revealed that the amino acid substitutions in families 2 and 182 4, both located within the DCC/Netrin-1 binding interface, are predicted to be most disruptive. 183 The p.(Val793Gly) substitution abolishes a hydrophobic interaction with Thr147 of Netrin-1 184 while p.(Gly805Glu) introduces a highly unfavorable charged moiety within a hydrophobic 185 pocket, disrupting interaction with Leu113 of Netrin-1 (Fig. S7-8). The predicted effects of the 186 three substitutions within the Netrin-1 binding region but outside the binding interface (Fig. S9-187 12) are consistent with *in vitro* studies demonstrating that even conservative mutations to residues in this binding region can disrupt DCC dimerization, Netrin-1 binding and axon guidance¹⁴. 188

189

190 In addition to the effect of sex hormones and the type and location of DCC mutations, 191 developmental differences between the CC and corticospinal tract may also contribute to the 192 variable ACC/MM phenotypes. Callosal and sub-cerebrally projecting pyramidal neurons of the 193 cortex are specified at early stages of development and the molecular identity of each population directly affects its axonal connectivity¹⁶. While corticospinal axons utilise DCC/Netrin-1 194 195 signalling to reach the midline, callosal axons use DCC/Netrin-1 chemoattraction to attenuate ROBO1/SLIT-2-mediated chemorepulsion to approach and cross the midline¹⁷. Therefore, a 196 197 DCC mutation may differentially affect commissural versus subcerebral axon trajectories, leading 198 to ACC, MM or both. MM were consistently associated with reduced crossing of descending 199 corticospinal tract projections at the pyramidal decussation in this study as well as in individuals with *RAD51*-related MM¹⁸, suggesting that *DCC*-mediated MM are primarily the result of 200 201 corticospinal tract decussation abnormalities.

202

203 In conclusion, our results provide compelling evidence that DCC mutations cause isolated ACC 204 in humans, in addition to the previously-reported MM phenotype. The factors determining the 205 phenotypic variability are complex and likely include the hormonal context during development, 206 the type and location of *DCC* mutation, and the genetic background of the individual. Although 207 the full spectrum of phenotypes associated with *DCC* mutations remains to be fully characterized, 208 individuals described in this study have an intellectual quotient within the normal/borderline 209 range. Heterozygous mutations in DCC therefore appear to result in isolated ACC with a mild 210 phenotype and favorable cognitive outcomes, contrasting with the unfavourable developmental 211 outcomes associated with syndromic ACC. Given the high frequency of DCC mutations detected 212 in our cohorts, this observation has prenatal diagnostic and parental counselling implications for 213 fetuses with ACC as the condition currently has unclear prognostication. Our data suggest that

- the prenatal detection of isolated ACC related to a pathogenic *DCC* mutation indicates a lower
- 215 risk of an abnormal neurodevelopmental outcome.

URLs. 1000 Genomes Project, http://www.1000genomes.org/; Exome Variant Server,
http://exac.broadinstitute.org/; SIFT, http://sift.jcvi.org/; PolyPhen-2,
http://genetics.bwh.harvard.edu/pph2/; dbSNP, https://www.ncbi.nlm.nih.gov/SNP/; PyMOL,
https://www.pymol.org/.

220

221 Data Availability.

Families included in this study have not consented to have Next Generation Sequence data publicly released. Variants identified in this study have been deposited into ClinVar for immediate release (SUB2184411) and accession numbers are pending final processing.

225

226 ACKNOWLEDGMENTS AND FUNDING

227 We thank the families and the Lefroy family for their participation in and support of this study. 228 We thank the DNA and cell bank of the ICM (Paris, France) for DNA extraction, Sinead Eyre 229 (QBI) for study co-ordination and Mike Kean (RCH) and Marc Seal (MCRI) for assistance with 230 MRI protocols and scanning. This work was funded in part by National Health and Medical 231 Research Council (NHMRC) Australia Project Grants (GNT1059666, GNT631466, 232 GNT1064174, GNT1048849, GNT1104455, GNT1064174), the Agence Nationale de la 233 Recherche (ANR Blanc CILAXCAL, ANR Blanc HARTaGeNe), Assistance Publique des 234 Hôpitaux de Paris (APHP), the "programme hospitalier de recherche clinique" (PHRC) 235 ACCREM, and the "Investissements d'Avenir" programme ANR-10-IAIHU-06 (IHU-A-ICM). 236 APLM and LM are supported by an Australian Postgraduate Award, TJE is supported by a 237 University of Queensland Research Scholarship and AP is supported by a QBI PhD scholarship. 238 SH and AQ are respectively supported by a master and a doctoral grant from the Fondation pour 239 la recherche médicale (FRM). MB is supported by an NHMRC Senior Research Fellowship and

an NHMRC Program Grant (GNT1054618). EHS is supported by a grant from the NIH,
2R01NS058721 and RJL is supported by a Melbourne Children's Clinician Scientist Fellowship.
LJR is supported by an NMHRC Principal Research Fellowship, and PJL is supported by a
NHMRC Career Development Fellowship (GNT1032364). CD and CN are members of the BioPsy Labex. This work has been supported in part by the Victorian Government's Operational
Infrastructure Support Program and Australian Government NHMRC IRIISS.

246

247 AUTHOR CONTRIBUTIONS

Ashley Marsh - formulation of theory and prediction, contributions to experimental conception and design, acquisition, analysis and/or interpretation of data and drafting the article and revising it critically for important intellectual content

Delphine Héron - contributions to experimental conception and design, interpretation of data and
 revising the article critically for important intellectual content

253 Timothy Edwards - formulation of theory and prediction, contributions to experimental 254 conception and design, acquisition, analysis and/or interpretation of data and drafting the article 255 and revising it critically for important intellectual content

256 Charles Galea - formulation of theory and prediction and acquisition, analysis and/or

257 interpretation of data

258 Angélique Quartier- acquisition, analysis and interpretation of data

259 Caroline Nava - contributions to experimental conception and design, acquisition, analysis and

260 interpretation of data, and revising the article critically for important intellectual content

261 Agnès Rastetter - acquisition, analysis and interpretation of data

262 Marie-Laure Moutard - acquisition, analysis and interpretation of data and revising the article

263 critically for important intellectual content

- 11 -

- 264 Vicki Anderson acquisition, analysis and/or interpretation of data and drafting the article or
- 265 revising it critically for important intellectual content
- 266 Pierre Bitoun acquisition, analysis and interpretation of data
- 267 Jens Bunt acquisition, analysis and/or interpretation of data
- 268 Anne Faudet acquisition, analysis and/or interpretation of data
- 269 Catherine Garel acquisition, analysis and/or interpretation of data
- 270 Greta Gillies acquisition, analysis and interpretation of data
- 271 Ilan Gobius acquisition, analysis and interpretation of data
- 272 Justine Guegan analysis of data
- 273 Solveig Heide acquisition, analysis and/or interpretation of data
- 274 Boris Keren acquisition, analysis and/or interpretation of data
- 275 Fabien Lesne acquisition, analysis and/or interpretation of data
- 276 Vesna Lukic acquisition, analysis and/or interpretation of data
- 277 Simone Mandelstam- acquisition, analysis and/or interpretation of data and drafting the article or
- 278 revising it critically for important intellectual content
- 279 George McGillivray acquisition, analysis and/or interpretation of data and drafting the article or
- 280 revising it critically for important intellectual content
- 281 Alissandra McIlroy acquisition, analysis and/or interpretation of data
- 282 Aurélie Meneret acquisition, analysis and/or interpretation of data
- 283 Cyril Mignot acquisition, analysis and/or interpretation of data, and revising the article critically
- 284 for important intellectual content
- 285 Laura Morcom acquisition, analysis and/or interpretation of data
- 286 Sylvie Odent acquisition, analysis and/or interpretation of data
- 287 Annalisa Paolino acquisition, analysis and/or interpretation of data

Kate Pope - acquisition, analysis and/or interpretation of data 288 289 Florence Riant - acquisition, analysis and/or interpretation of data 290 Gail Robinson - acquisition, analysis and/or interpretation of data 291 Megan Spencer-Smith - acquisition, analysis and/or interpretation of data 292 Myriam Srour - acquisition, analysis and/or interpretation of data 293 Sarah Stephenson - contributions to experimental conception and design 294 Rick Tankard - acquisition, analysis and/or interpretation of data 295 Oriane Trouillard - acquisition, analysis and/or interpretation of data 296 Quentin Welniarz - acquisition, analysis and/or interpretation of data 297 Amanda Wood - acquisition, analysis and/or interpretation of data 298 Alexis Brice - acquisition, analysis and/or interpretation of data and revising the article critically 299 for important intellectual content 300 Guy Rouleau - acquisition, analysis and/or interpretation of data and revising the article critically 301 for important intellectual content 302 Tania Attié-Bitach- contributions to experimental design, and revising the article critically for 303 important intellectual content 304 Martin Delatycki - drafting the article or revising it critically for important intellectual content 305 Jean Louis Mandel - contributions to experimental conception and design, interpretation of data 306 and revising the article critically for important intellectual content 307 David Amor - drafting the article or revising it critically for important intellectual content 308 Emmanuel Roze - acquisition, analysis and/or interpretation of data and revising the article 309 critically for important intellectual content 310 Amélie Piton - contributions to experimental conception and design, interpretation of data and 311 revising the article critically for important intellectual content

Melanie Bahlo - acquisition, analysis and/or interpretation of data and drafting the article or
 revising it critically for important intellectual content

314 Thierry Billette de Villemeur - acquisition, analysis and/or interpretation of data and revising the

315 article critically for important intellectual content

316 Elliott Sherr - formulation of theory and prediction, acquisition, analysis and/or interpretation of

317 data and drafting the article and revising it critically for important intellectual content

Richard Leventer - formulation of theory and prediction, acquisition, analysis and/or
interpretation of data and drafting the article and revising it critically for important intellectual

320 content

321 Linda Richards - formulation of theory and prediction, contributions to experimental conception

322 and design, acquisition, analysis and/or interpretation of data and drafting the article and revising

323 it critically for important intellectual content

324 Paul Lockhart - formulation of theory and prediction, contributions to experimental conception

325 and design, acquisition, analysis and/or interpretation of data and drafting the article and revising

326 it critically for important intellectual content

327 Christel Depienne - formulation of theory and prediction, contributions to experimental 328 conception and design, acquisition, analysis and/or interpretation of data and drafting the article 329 and revising it critically for important intellectual content

330

331 COMPETING FINANCIAL INTERESTS

332 The authors declare no competing financial interests.

333 **REFERENCES**

- 1. Lindwall, C., Fothergill, T. & Richards, L.J. Curr Opin Neurobiol 17, 3-14 (2007).
- 335 2. Chedotal, A. *Curr Opin Neurobiol* **21**, 68-75 (2011).
- 336 3. Paul, L.K. *et al. Nat Rev Neurosci* **8**, 287-99 (2007).
- 337 4. Glass, H.C., Shaw, G.M., Ma, C. & Sherr, E.H. Am J Med Genet A 146a, 2495-500
 338 (2008).
- 339 5. Rouleau, C. et al. Arch Dis Child Fetal Neonatal Ed 96, F360-4 (2011).
- 6. Edwards, T.J., Sherr, E.H., Barkovich, A.J. & Richards, L.J. Brain 137, 1579-613 (2014).
- 341 7. Sotiriadis, A. & Makrydimas, G. *Am J Obstet Gynecol* **206**, 337.e1-5 (2012).
- 342 8. Fazeli, A. et al. Nature **386**, 796-804 (1997).
- 343 9. Srour, M. et al. Science **328**, 592 (2010).
- 344 10. Meneret, A. *et al. Neurology* **82**, 1999-2002 (2014).
- 345 11. Ardekani, B.A., Figarsky, K. & Sidtis, J.J. Cereb Cortex 23, 2514-20 (2013).
- 346 12. Moffat, S.D., Hampson, E., Wickett, J.C., Vernon, P.A. & Lee, D.H. *Brain Res* 767, 297304 (1997).
- 348 13. Chura, L.R. et al. Psychoneuroendocrinology **35**, 122-32 (2010).
- 349 14. Finci, L.I. et al. Neuron 83, 839-49 (2014).
- 350 15. Xu, K. et al. Science **344**, 1275-9 (2014).
- 16. Greig, L.C., Woodworth, M.B., Galazo, M.J., Padmanabhan, H. & Macklis, J.D. *Nat Rev*
- 352 *Neurosci* **14**, 755-69 (2013).
- 353 17. Fothergill, T. et al. Cereb Cortex 24, 1138-51 (2014).
- 354 18. Gallea, C. *et al. Brain* **136**, 3333-46 (2013).

355 Figure legend

Figure 1 *DCC* mutations cause isolated ACC and/or MM associated with significantly reduced
 crossing of descending corticospinal tract projections at the pyramidal decussation.

a. All available family samples were analysed; m=mutation; black dot=mutation carrier;
blue=partial ACC; black=complete ACC and grey=MM. * indicates neuroimaging data for
mutation carrier or individual with MM.

b. Midsagittal MRI of control and family proband/representative individual (1-2=complete ACC;
3=near complete ACC with thin rostrum and genu remaining and 4=partial ACC with absence of
the rostrum and genu).

364 c. Group-wise comparison of laterality coefficient in both families (family 2, n=4; family 4, n=3) 365 were compared to controls (n=6). For each individual, a laterality coefficient for the corticospinal 366 tract was calculated as the ratio of the difference between the numbers of crossed and uncrossed 367 streamlines to the total number of streamlines. Right and left coefficients were averaged to find 368 the laterality coefficient of each individual. Greater positive values indicate more crossed and 369 negative values more uncrossed streamlines (mean +/- S.D, * p=0.0238 ; ** p=0.0095; two-tailed 370 Mann-Whitney U-test).

d. Protein domain structure depicting the location of the DCC truncation (red square and triangle)
and missense mutations (colored dots). The Netrin-1 binding region is indicated, IgC2,
immunoglobulin-like type C2 domain; FN3, fibronectin type III-like domain; TM,
transmembrane domain; P1-3, conserved motifs.

- 375 Table 1: Summary of DCC mutations identified in individuals with ACC (+/-MM) in this study. cACC, complete isolated agenesis of
- 376 the corpus callosum; pACC, partial isolated agenesis of the corpus callosum; MM, mirror movements; IgC2, immunoglobulin-like type
- 377 C2 domain; FN3, fibronectin type III-like domain; ExAC, Exome Aggregation Consortium; dbSNP, dbSNP reference SNP
- 378 identification number. Reference sequences used are NM_005215.3 and NP_005206.2.

Family number	Phenotype	cDNA	Protein	Protein domain	SIFT	PolyPhen-2	ExAC	dbSNP
1	cACC or pACC ± MM	c.925delA	p.(Thr309ProfsTer26)	IgC2-3	-	-	No	-
2	cACC & MM	c.2378T>G	p.(Val793Gly)	FN3-4	Deleterious	Probably damaging	No	-
3	cACC or MM ± pACC	c.823C>T	p.(Arg275Ter)	IgC2-3	-	-	No	-
4	pACC &/or MM	c.2414G>A	p.(Gly805Glu)	FN3-4	Deleterious	Probably damaging	No	-
5	cACC	c.1790G>C	p.(Arg597Pro)	FN3-2	Deleterious	Probably damaging	No	-
6	cACC	c.2227A>T	p.(Met743Leu)	FN3-4	Deleterious	Benign	No	rs199651452
7	cACC	c.2260G>A	p.(Val754Met)	FN3-4	Deleterious	Possibly damaging	Yes (x19)	-
8	cACC	c.2677G>A	p.(Ala893Thr)	FN3-5	Deleterious	Benign	No	-
9	cACC	c.3649A>G; c.3748G>A	p.(Met1217Val); p.(Ala1250Thr)	Cytoplasmic	Tolerated; Tolerated	Benign; Probably damaging	No; Yes (x2)	

