

Bioactive Organic-Inorganic Hybrids with Improved Mechanical Performance

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Abstract: New sol-gel functionalized poly-ethylene glycol (PEGM)/SiO₂-CaO hybrids were prepared with interpenetrating networks of silica and PEGM through the formation of Si-O-Si bonds. Bioactive and mechanical properties were investigated for a series of hybrids containing varying organic/inorganic ratios and PEG molecular weights. In contrast to the unmodified PEG/SiO₂-CaO hybrids, which rapidly dissolved and crumbled, the epoxy modified hybrids exhibited good mechanical properties and bioactivity. The compressive strength and Young's modulus were greater for higher molecular weight PEGM hybrids (PEGM600 compared to PEGM300). Compressive strengths of 138 MPa and 81 MPa were found for the 50:50 and 60:40 organic/inorganic hybrid samples respectively, which are comparable with cortical bone. Young's Modulus values of ~ 800 MPa were obtained for the 50:50 and 60:40 organic/inorganic hybrids. Bioactivity tests were conducted by immersing the hybrids into simulated body fluid and observing the formation of apatite. Apatite formation was observed within 24 hours of immersion. PEGM600 hybrids showed enhanced apatite formation compared to PEGM300 hybrids. Increased apatite formation was observed with increasing organic/inorganic ratio. 70:30 and 60:40 hybrids exhibited the greatest apatite formation. All PEGM hybrids samples had good cell viability and proliferation. The 60:40 PEGM600 hybrids displayed the optimal combination of bioactivity and mechanical strength. The bioactivity of these hybrids, combined with the enhanced mechanical properties, demonstrate that these materials have significant potential for bone regeneration applications.

Keywords: hybrid, PEG, bioactivity, bone regeneration

1 Introduction

Bone is the most widely transplanted material, with the exception of blood only [1]. Each year more than 2.2 million bone graft operations are performed to repair bone defects in orthopaedics and

dentistry [2, 3]. Bioactive glasses represent one of the most promising synthetic bone replacement materials; these materials have the potential to be more bioactive than calcium phosphate-based materials. Bioactive glasses, developed by Larry Hench in 1969 show class A bioactivity; they bond to bone and stimulate new bone growth even away from the glass–bone interface[4, 5].

Melt-derived Bioglass[®] (46.1% SiO₂, 24.4% Na₂O, 26.9% CaO, and 2.6% P₂O₅ mol%) is used commercially in products such as Novabone[®] and NovaMin[®] for repairing bone defects and re-mineralizing dental enamel[5, 6]. Under physiological conditions the glass slowly dissolves releasing controlled amounts of Ca, P and Si. The soluble Ca and P then precipitate onto the surface to form an amorphous calcium phosphate layer which then crystallizes into apatite (the mineral component of bone)[7]. Studies have shown that the controlled release of Ca, P and Si can stimulate gene transcription in osteoblasts [8, 9]. The glass surface also supports cell adhesion and proliferation. There are however limitations; Bioglass[®] requires very high processing temperatures (~1400 °C [10]), is difficult to process into scaffolds, fibres or coatings, and has a very small processing window which can result in crystallization. In addition, melt-quench glasses have low surface areas and low porosity which is not ideal for cell ingrowth or vascularization. Sol-gel derived bioactive glasses have been developed to overcome many of these obstacles[11]. Sol–gels, prepared using a low-temperature hydrolysis and condensation process, have many advantages compared with their melt–quench derivatives [12-14]. The sol-gels process has greater versatility; bioactive particles, foams, coatings and fibres can all be prepared using this methodology. The resultant gels have a nanometer-scale textural porosity, high specific surface area and abundant exposed silanol groups on surface [15-17]. Apatite formation occurs in the macropores resulting in a much greater degree of apatite growth [18, 19]. Despite their excellent bioactivity and acceptable compressive strength their poor tensile strength and brittle nature has severely limited their applications.

Sol-gel derived glasses can be combined with organic polymers to improve their mechanical strength and toughness[20, 21]. Bioactive glass particles can be dispersed in a polymer matrix to form composites[22]. However, the bioactive particles are generally covered by the polymer matrix and are therefore not in direct contact with body fluid before polymer degradation, resulting in slow bonding to bone tissue *in vivo*[23]. In addition, the bioactive glass particles are usually on the micrometre-scale, thus likely to cause inhomogeneity. Alternatively, inorganic / organic hybrids can be synthesised by introducing the polymer at an early stage e.g. around the hydrolysis stage. Organic-inorganic hybrids show similar improvements in mechanical properties compared to the composites but also show much better bioactivity due to the mixing of glass and polymer on molecular scale. Therefore, hybrids have attracted much attention owing to these advantages.

Hybrids can be classified as class I and class II hybrids according to the interaction between inorganic chains and organic chains. In class I hybrids inorganic chains entangle with organic chains with only weak (hydrogen and/or van der Waals bonding) interactions, whereas by modifying the polymer to react with the inorganic phase strong covalent bonds can be formed (class II hybrids) [24]. Class II hybrids have previously been fabricated using alkoxy silane functionalized

polymer as precursors in the form of scaffolds, electrospun fibres or monoliths[25-27]. These hybrids have been shown to exhibit mechanical properties analogous to human cancellous bones[28-30]. For example Salinal *et al.*[31] have prepared dense and homogeneous crack-free monoliths of PDMS-modified CaO-SiO₂ system. These hybrids are silicate networks covalently bonded to PDMS which show apatite formation when immersed in simulated body fluid (SBF). It was also shown that their elastic modulus decreased with PDMS content but their apatite forming ability increased with calcium content[32]. Biodegradable Silica/poly(ϵ -caprolactone) (PCL) have also been prepared, in which hydroxyl groups at either end of the PCL polymer chains were modified with 3-isocyanatopropyl triethoxysilane (IPTS), resulting in a polymer end capped with a triethoxysilyl group. The end capped PCL then was introduced into a sol to synthesize bioactive glass/PCL hybrids[33, 34]. Bioactive silica-poly(γ -glutamic acid) hybrids have also been obtained by covalently bonding the γ -PGA and bioactive silica through an organosilane coupling agent, glycidoxypropyl trimethoxysilane (GPTMS)[24], and calcium was confirmed to be incorporated in the silica network by using calcium alkoxide precursor. However, sometimes the resulting hybrids are not sufficiently hydrophilic for ion exchange with the surrounding physiological fluids. Hydrophilic organic components have been introduced to prepare organic-inorganic hybrids. For example, natural polymer gelatin was functionalized using GPTMS, which was then introduced into a sol of hydrolyzed TEOS to covalently link gelatin to silica with -Si-O-Si-. Then the wet gels were freeze dried to form porous scaffolds, the mechanical properties of which are similar to the cancellous bones [35]. Granqvist *et al.* have developed PEG-siloxane fibres by functionalising PEG end groups with 3-Isocyanatopropyl triethoxysilane[25]. The functionalised PEG was incorporated into the siloxane network through covalent bonds resulting in fibres with much higher elasticity compared with non-functionalised fibres. The fibres were shown to be effective substrates for the nucleation and growth of bone-like apatite.

Traditionally calcium has always been incorporated in sol-gels in the form of calcium nitrate which then requires the sol-gel to be heated >400°C to burn off the toxic nitrates and for calcium to be incorporated into the silicate network [19, 36]. Nitrates are therefore unsuitable for hybrid materials containing polymers which decompose at high temperatures. However, it has recently been shown that the precursor calcium methoxyethoxide (CME) is suitable introducing calcium into hybrids[36, 37]. Calcium has been shown to enter the silicate network at processing temperatures as low as 60°C when using CME.

The purpose of the present study was to prepare homogeneous organic-inorganic hybrid materials for bone repair using low molecular weight (300, 600) functionalized PEG. The inorganic phase consisted of (SiO₂)₇₀-(CaO)₃₀ (S70C30, the archetypal bioactive sol-gel). The organic phase consisted of polyethylene glycol (PEG), a biocompatible polymer (and bioresorbable for low molecular weight polymers[38]) which is completely soluble in both water and ethanol, easy to modify and can be incorporated into a typical sols for sol-gel hybrid synthesis. PEG ends were first modified with epoxy ethane, then ammoniaprolyltriethoxysilane was added to react with epoxy ethane to form PEG end capped siloxane (PEGM). The organic-inorganic hybrid materials were prepared by introducing PEGM into the sol-gel process, using TEOS and calcium methoxyethoxide

as precursors, so that the silica network and organic polymer form interpenetrating networks at the nanoscale. The aim was to investigate whether this system can improve the mechanical properties of the hybrid materials whilst retaining the apatite forming bioactivity of the inorganic sol-gel phase.

2 Materials and Methods

2.1 Materials and Sample Preparation

The following precursors were used in the sol-gel preparation: PEG300 (285-315), PEG600 (585-615), ammoniapropyltriethoxysilane (97%) and methoxyethanol (99.8%), (3-Aminopropyl)triethoxysilane (APTES) (97%) (Sigma Aldrich), Epichlorohydrin, metallic calcium and tetraethyl orthosilicate (TEOS, $\geq 99.0\%$) (Sinopharm Chemical Reagent Co., Ltd).

The preparation was carried out in a 4 step process as shown in Figure 1.

Calcium methoxyethoxide preparation: One gram of Ca metal was added into 24mL of methoxyethanol under reflux for 24h at 80°C [39]. Calcium methoxyethoxide solution was obtained from the supernatant fluid after centrifuging (Ca-methoxyethoxide, 17% in methoxyethanol).

Modifying PEG with EO (PEG-EO): PEG (50mmol) was dissolved in 50mL of tetrahydrofuran. Epichlorohydrin (200mmol) and NaH (2.1 g, 60 % in mineral oil) were added and the medium was stirred under reflux (66°C) for 24h. After the solution cooled to room temperature, 30% H₂SO₄ in methanol was added to neutralize the solution. The solvent and excess epichlorohydrin was then removed under reduced pressure. ¹H NMR spectra were recorded at 25°C on an Avance III 400 spectrometer to confirm the EO modification. Chemical shifts are reported in parts per million (ppm) and a reference standard of CDCl₃ (7.26ppm) was used. FTIR spectra were recorded for PEG, PEG-EO and PEGM using a Bruker Equinox 55 spectrometer to monitor and confirm the reaction process.

Modification of PEG-EO with siloxane (PEGM): PEG300-EO (4.14g, 0.01mol) was dissolved into 50ml of CH₂Cl₂, 4.68ml (0.02mol) ammonia propyl triethoxysilicon (APTES) (KH550) was then added and stirred for 24h at room temperature. The solvent was removed at reduced pressure. This method was replicated for the PEG600-EO (7.14g, 0.01mol) sample.

Preparation of organic-inorganic hybrid material: under constant stirring PEGM were firstly mixed with TEOS, then Ca-methoxyethoxide was added at different organic/inorganic molar ratios of 70:30, 60:40, 50:50 and 40:60 whilst maintaining the inorganic composition of S70C30. After 30min, water (H₂O/TEOS=2, molar ratio) was added drop wise into the solution and the solution gels within an hour. The resultant gels were aged at ambient temperature for two days then dried in an oven, firstly at 40°C for two weeks and then at 120°C for a further week. Finally the samples were dried at 280°C for 1 h to ensure full condensation and complete evaporation of the solvent. A control sample of PEG600 with a 60:40 organic: inorganic ratio was prepared using unmodified PEG to act as a reference. ²⁹Si solid state MAS NMR spectroscopy experiments were carried out on the dried materials using a Bruker Avance III 400 MHz instrument providing ²⁹Si Larmor frequencies of 79.30 MHz. 4 mm MAS BB probes spinning at 12 kHz were used. A 2.5 μs pulse

($\sim 60^\circ$ tip angle) was used, and the recycle delay was 90 s. ^{29}Si NMR spectra were referenced using tetramethylsilane ($\delta_{\text{Si}} = 0$ ppm).

2.2. Bioactivity testing

Bioactivity was tested *in vitro* by immersing 150 mg monoliths in 100 ml of simulated body fluid at $36.5 \pm 0.5^\circ\text{C}$. SBF was prepared by dissolving reagents NaCl, NaHCO_3 , KCl, $\text{K}_2\text{HPO}_4 \cdot 3\text{H}_2\text{O}$, $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$, CaCl_2 , and Na_2SO_4 in deionized water[40]. The solution was buffered to pH 7.4 with Tris-(hydroxymethyl)-aminomethane ($((\text{CH}_2\text{OH})_3\text{CNH}_2)$) and hydrochloric acid to give a resultant ion concentration similar to human plasma. After immersing in SBF for different times the samples were washed gently with pure water and left to dry in a desiccator without heating. Apatite formation was evaluated using scanning electron microscopy (SEM), Energy Disperse X-Ray Spectroscopy (EDXS), Fourier transform infrared (FTIR) and X-ray diffraction (XRD).

SEM / EDXS: Following immersion the samples were dried and then coated with a thin layer of gold (Au) by sputtering (SCD 500). The surface development was observed using a JEOL-6700 instrument operating at a voltage of 15 kV.

XRD: Following immersion the samples were dried and then finely powdered. X-ray diffraction spectra were collected using a Rigaku (D/MAX 2500) instrument with $\text{Cu K}\alpha$ radiation ($\lambda = 1.54\text{\AA}$), operated at 40 kV and 200 mA. Data was collected for 2θ values between 10° to 60° with a step size of 0.02° .

FTIR: Fourier transform infrared spectra were collected in the range $2000\text{--}400\text{ cm}^{-1}$ on a Bruker Equinox 55 instrument. All samples were diluted with dry KBr, ground down to fine powders and pressed into pellets. Measurements were made at ambient temperature.

2.3 Mechanical testing

Compressive mechanical properties of the specimens were evaluated using general a purpose-testing machine (Instron3365, Instron Co., Canton, MA, USA) at $23 \pm 2^\circ\text{C}$ and $50 \pm 5\%$ relative humidity conditions. Specimens were cut from homogeneous regions of the organic-inorganic hybrids to form blocks measuring $4\text{mm} \times 4\text{mm} \times 8\text{mm}$. These samples were positioned between parallel plates and compressed with a crosshead speed of 0.5mm min^{-1} and a 5.0 kN load cell. A minimum of five repeats were taken per sample to provide an average. The mean and standard deviation were reported. The Young's modulus was determined from the slope of the initial linear elastic portion of the stress-strain curve. Compressive strength was taken as the maximum strength of the sample until failure and elongation to failure was the percentage strain at fracture.

2.4 Cell evaluation

Preparation of hybrids extracts: Organic/inorganic hybrids were soaked in α -MEM, preheated to 37°C , extracting vehicle ratio was $3\text{cm}^2/\text{ml}$, as specified by ISO 10993-12:2007. Organic/inorganic hybrids, which were previously sterilized under UV light for 12 h. After 1, 3 and 7 days at 37°C under sterile conditions, α -MEM was filtered to eliminate hybrids, and these extracts (E1, E2 and

E3, respectively) were used as culture medium after adding 10% fetal bovine serum (FBS).

Cell culture: A preosteoblast cell line (MC3T3-E1; ATCC, CRL-2593, Rockville, MD, USA) was used to investigate the cell compatibility *in vitro*. The MC3T3-E1 cells were cultured in a humidified incubator under an atmosphere containing 5% CO₂ at 37°C. Dulbecco's Modified Eagle Medium (α -MEM) supplemented with fetal bovine serum (FBS, 10%), penicillin (100 μ g/ml) and streptomycin (100 μ g/ml) was used as the culture medium. Cells were subcultured every 5 days and maintained at 37°C in a humidified incubator with 5% CO₂.

Cell viability assays: A Cell Count Kit-8 (CCK-8, Beyotime, Jiangsu, China) was employed to quantitatively evaluate cell viability [41]. MC3T3-E1 was seeded on 96-well culture plates at a density of 1.7×10^4 cells/ml, after cell adhesion was verified; the culture medium was replaced by different extracts (E1, E2 and E3) of the organic/inorganic hybrids. The cells incubated in α -MEM without organic/inorganic hybrids extract were used as a control. The cells were incubated in the culture plates for 3 and 6 days. Then, the CCK-8 solution (20 μ l per well) was added and incubated for 4h at 5% CO₂ and 37°C, 100 μ l of the reacted reagent from each well was transferred to 96-well plates, and the absorbance at 450 nm was determined using a micro-plate spectrophotometer (MD, SpectraMax M2, USA). Six parallel replicates of each sample at each time point were prepared so that statistics could be performed.

2.5 Degradation studies

Hybrids (75 mg) in triplicates were immersed in 50 mL of SBF at $36.5 \pm 0.5^\circ\text{C}$ and stirred at 120 rpm. At various time points (1, 4, 24, 72, 168, 336, 504 and 672h), the hybrids were removed from their respective containers and dried under vacuum prior to weighing. To obtain the rate of weight loss, the initial weight (M_0) of each sample was measured as well as the weight loss at time t (M_t) to give a weight loss% : $\text{weight loss\%} = (M_0 - M_t) / M_0 * 100\%$. pH values of the extracts were recorded after 1, 3 and 6 days immersion.

2.6 Statistical analysis

The one-way analysis of variance (ANOVA) was used to evaluate significant differences between means in the measured data. Each experiment was repeated three times. All quantitative data are presented as mean \pm standard deviation.

3 Results

3.1. Sample characterisation

PEG end groups were successfully modified with epoxy (EO) as shown in Figure 2. The ¹H NMR spectra confirms the addition of the epoxy functional groups. The spectra are assigned as follows: ¹H NMR (400MHz, CDCl₃) δ (ppm), 3.58 (m, 1H, H_e), 3.43(m, 28H, H_f), 3.19(m, 1H, H_d), 2.92(m, 1H, H_c), 2.56(m, 1H, H_b), 2.38(m, 1H, H_a)[42]. The reaction process of PEG-EO to PEGM and corresponding FTIR spectra are shown in Figure 3. A feature at 914 cm^{-1} appears for PEG-EO which is consistent with an epoxy group[43, 44]. The epoxy peak then disappears for the PEGM as

expected (see Figure 3a bottom). Clear amorphous monoliths were produced as shown in Figure 4. The orange colour emanates from the CME precursor which is a dark red/brown colour as previously reported.

3.2. Structure characterization

Figure 5 shows the ^{29}Si MAS NMR spectra of (a) PEGM300/TEOS (calcium free) (b) the PEGM/TEOS/CME hybrid and (c) the unmodified PEG300/TEOS/CME hybrid. The modified PEG hybrids (samples a and b) both show the presence of T^n and Q^n species (T^n and Q^n correspond to the structure of $\text{CSi}(\text{OSi})_n(\text{R})_{3-n}$ and $\text{Si}(\text{OSi})_n(\text{R})_{4-n}$ respectively, where R is a non-bridging oxygen). In contrast the unmodified PEG hybrid (sample c) only showed the presence of Q^n species. Results of the Gaussian fitting of T^n and Q^n distributions of the ^{29}Si MAS NMR spectra are summarised in Table 1. The ratio of T^3/T^{tot} and Q^4/Q^{tot} in Table 1 show the degree of condensation of the PEGM and silica network respectively. For the TEOS-derived silica network, ^{29}Si exhibit several overlapping contributions associated with Q^n units: three resonances at ~ 95 , ~ 101 and ~ 110 ppm were assigned to Q^2 , Q^3 and Q^4 units respectively. In addition to these Q^n resonances, the ^{29}Si MAS NMR spectra of sample a and b showed two peaks at ~ 60 and ~ 65 ppm characteristic of T^2 and T^3 units resulting from the hydrolysis/condensation of the terminal triethoxysilane groups of PEGM. Despite poly-condensation between PEGM chains not being excluded, the high occurrence of T^3 units suggests that PEGM is successfully grafted to the BG network, thereby coupling the organic and inorganic parts of the hybrid to form a Class II hybrid. Sample a had higher T^3/T^{tot} and Q^4/Q^{tot} values compared to samples b and c, which suggests that the silica species were highly condensed.

3.3. Bioactivity

Apatite formation was observed on the monolith's surface after immersion in SBF. Figure 6 illustrates a representative SEM image showing both hemispherical and needle like apatite formation. Needle like formations are consistent with previous reports of apatite formation on bioactive sol-gels[45]. Apatite formation occurs within 24 hours of immersion and become denser with increasing time.

X-ray diffraction results are shown in Figures 7-9. Figure 7 illustrates the formation of apatite as a function of SBF immersion time for PEGM600 with an organic/inorganic molar ratio of 60:40. HA diffraction peaks appear after only 24 hours and then increase in magnitude after 3 and 7 days confirming apatite formation on the surface of these materials. Monoclinic HA is shown for reference standard[46]. Figures 8 and 9 show the apatite diffraction peaks as a function of organic/inorganic ratio for the PEGM600 and PEGM300 hybrids respectively.

The intensity of the apatite diffraction peaks increases with increasing organic/inorganic ratio for both PEGM600 and PEGM300 hybrids. It is evident that the 60:40 and 70:30 hybrids show the greatest bioactivity (apatite formation). For all organic/inorganic ratios the PEGM600 spectra shows significantly more apatite formation demonstrating that PEGM600 hybrids are more bioactive compared to their equivalent PEGM300 counterparts.

EDXS results are shown in Figure 10 as a function of SBF immersion time. Prior to immersing in SBF, the hybrid surface consisted of Si, C and O as expected. Within a day of immersion in SBF the hybrid surface is rapidly covered with a calcium phosphate layer as expected, in addition the Si and C signal from the bulk hybrid is significantly attenuated by the Ca/P surface formation. The Ca and P signals continue to increase with time (3 and 7 day immersion) and the Si signal is further attenuated.

FTIR spectra are shown in Figure 11. The pronounced band $\sim 1090\text{ cm}^{-1}$ and the smaller peak $\sim 790\text{ cm}^{-1}$ are attributed to Si-O-Si vibrations [47]. The broad band $\sim 3440\text{ cm}^{-1}$ and the sharper peak $\sim 1630\text{ cm}^{-1}$ are attributed to O-H stretching [47]. Peaks at ~ 459 , 567 and 607 cm^{-1} are attributed to phosphate absorption bands [45]. These peaks evolve as a function of immersion time in SBF further supporting the formation of phosphates on the hybrids surface.

3.4. Mechanical Testing

Compressive mechanical testing was performed on the dried hybrid monoliths to test the strength and flexibility of the hybrids. Figure 12 shows the stress-strain behaviour and effect of organic/inorganic ratio on Young Modulus of hybrids, and Table 2 presents the corresponding Young's modulus and compressive stress data for PEGM300 and PEGM600 samples as a function of organic/inorganic ratio. The elongation to failure was greatly increased to 14-35% for the hybrids materials compared to 4.2% for sol-gel glasses (70S30C monoliths, sintered at 800°C)[48], indicating improved strain to failure and therefore increased toughness. Two clear trends are also observed; firstly the mechanical properties increase with increasing inorganic ratio. The Young's modulus and compressive strength are greatest for the 50:50 hybrid with the overall trend 50:50 > 60:40 > 70:30 and this trend is observed for both the PEGM300 and PEGM600 hybrids. The second point is that Young's modulus and compressive stress are greater for the PEGM600 hybrid compared to its equivalent PEGM300 hybrid (for the same organic/inorganic ratio). The PEGM600 hybrid with a 50:50 organic/inorganic ratio was therefore found to have the best mechanical properties with 60:40 PEGM600 having very similar Young's modulus values. The unmodified PEG hybrid had very poor mechanical properties and easily crumbled, it was therefore not possible to measure the modulus or strength of these samples. PEG rapidly dissolved from the unmodified composite resulting in just the inorganic phases. Strong covalent links between the organic and inorganic chains are therefore essential in order to obtain tough composites which degrade uniformly acting as a single phase material.

3.5. Cell viability test

As shown in Figure 13, good cell proliferation (96%-157%) was observed in the presence of the three extracts (E1, E2 and E3). Good cell viability was observed for all the extracts, no significant difference in viability was observed for the different extracts after 3 days. However after 6 days the cell viability was higher for the both hybrids compared to the control. The cell proliferation for the PEGM600 hybrids extracts is slightly higher than PEGM300 hybrids (Figure 13a). The cell proliferation increased with culture time in each group (Figure 13b).

3.6. Degradation studies

A preliminary evaluation of the degradation behaviour of the hybrids was conducted through mass loss upon immersion in SBF solution. Results are summarised in Figure 14. Mass loss increased rapidly for both samples at during the first week before plateauing at ~ 43% at 3 weeks. These results are favourable compared to other organic/inorganic hybrids. For example, Russo *et al* report mass losses as high as 76% at 3 weeks for dPEG:GPTMS with a 1:2 ratio and losses of 52% at 3 weeks for dPEG:GPTMS with a 1:4 ratio [49].

Table 3 presents the pH data for the PEGM600 60:40 and PEGM300 60:40 sample as a function of immersion time. It can be seen that the pH value of the extracts are stable with only a small increase in pH. The pH of these hybrids are lower than values reported for NovaBone (7.64) and S70C30 scaffolds (8.19) after immersion for 72h [50]. It has been reported that higher pH value may be detrimental for cell growth. The PEG modified hybrids exhibit lower pH values compared to the same inorganic composition (S70C30) this indicates that PEG moieties may help stabilize the inorganic phase of the hybrid.

4 Discussion

PEG was successfully modified, via an epoxide functionalization, to yield PEGM/70S30C bioactive hybrids with covalent (Si-O-Si) bonds linking the organic and inorganic phases. A series of hybrids containing different organic/inorganic molar ratios and PEG molecular weights were prepared and characterised. In all cases the modified PEGM hybrids showed significantly improved mechanical properties compared to the unmodified PEG hybrids. The improvement in mechanical properties was attributed directly to the covalent bonding between the organic and inorganic phases for the modified PEG hybrids. The present samples exhibited favourable bioactivity and mechanical properties compared to equivalent materials previously reported in the literature. In previously studies, Liu *et al* prepared PEG/SiO₂-CaO-P₂O₅ hybrid xerogels by introducing PEG during the hydrolysis process to provide a molecular scale interaction between the organic and inorganic phases. The PEG molecular chains were found to penetrate into the SiO₂ network forming a semi-IPN structure. Since there was no covalent bonding between inorganic and organic phases, although a significantly increase in both the Young's modulus (320 to 420 MPa) and hardness was achieved compared to the pure inorganic material[47], it still could not reach the level achieved in the present study (in the present study we are achieving Young's modulus values in excess of 800 MPa for PEGM600 50:50 and 60:40 hybrids). Wang *et al* have developed nanoparticle-gelatin composites with enhanced mechanical properties [51]; the maximum value of Young's Modulus reported for these systems was 600 MPa compared to over 800 MPa in the current system.

Rhee has developed poly(ϵ -caprolactone)/silica hybrids for bone repair applications, with covalent bonding between inorganic and organic phases[33]. These materials show good apatite forming ability and Young's moduli of 200 – 600 MPa. Russo *et al* have recently developed silica/bis(3-aminopropyl) polyethylene glycol(dPEG) inorganic/organic hybrids by using 3-glycidopropyltrimethoxysilane (GPTMS) as a coupling agent to provide the covalent links[49]. The strain to failure increased from 4.2% for the sol-gel glass to 8-15% for the hybrids and

compressive strengths of 15-20MPa were reported which is comparable to cancellous bone (4-12 MPa) but low compared to cortical bone (100-230MPa). In the present study we are achieving compressive strength values of 138 and 81 MPa for PEGM600 50:50 and 60:40 hybrids respectively. In addition to the covalent bonding, the improvement in mechanical properties achieved in our study might be partly due to the choice of molecular weight of polymer segments used. Previous studies have employed polymers with molecular weights greater higher than 1,000[33, 49]. However, the polymer phase is mechanically weaker in compression compared to the inorganic phase. In the present study, PEG600 was found to be better than PEG300 suggesting there is also a lower limit for optimised polymer weight. However, we also note that it was not possible to form monoliths from higher molecular weight PEG (2000, 3000, 4000 and 6000). Therefore, an optimal molecular weight of polymer must to be chosen, in order to maximise the toughness of these hybrids.

Apatite formation was observed for 50:50, 60:40 and 70:30 PEGM600 and 60:40 and 70:30 PEGM300 hybrids. For a given organic/inorganic ratio the apatite formation of PEGM600 hybrids were consistently higher than the equivalent PEGM300 hybrids; this is attributed to the higher the molecular weight of the polymer. Apatite formation increased with increasing organic content with 60:40 and 70:30 (organic/inorganic molar ratio) showing the most promising formation. 50:50 and 40:60 hybrids showed minimal apatite formation. PEG is hydrophilic and relatively flexible, the increase in apatite formation with PEG molecular weight and PEG content are therefore likely to have resulted from better penetration of SBF into the material. This would enable the simulated fluid to react more quickly with the bioactive inorganic phases.

The 60:40 PEGM600 hybrids were the most promising materials demonstrating both good apatite formation and good mechanical properties. Young's Modulus values ~ 800 MPa are higher than those of cancellous bone and significantly higher than non-functionalised PEG hybrids and other values reported in the literature. Compressive strength of ~ 80 MPa is similar to values required for compact bone (100 MPa). The excellent biocompatibility and aqueous solubility of PEG molecules, as well as the bioactivity of the inorganic components, coupled with the improved mechanical properties mean these hybrids exhibit great potential for bone regeneration applications.

The hybrids demonstrate good biocompatibility for both cell viability and proliferation. The hybrids exhibit an initial rapid weight loss, which provides the essential release of Ca and Si that is necessary for apatite and to stimulate cellular activity [8, 52]. After the initial weight loss observed during the first week which peaked at 37% the weight-loss plateaus and only a further 4% weight-loss is observed during the subsequent 3 weeks. Apatite formation counteracts further weight-loss after the first week such that the total weight loss after 4 weeks is only 41%.

No significant difference was observed between the PEGM300 and PEGM600 samples. This confirms that the molecular weight has little effect on the degradation of hybrids, and the degradability may be controlled by inorganic phase. During the first week, the mass loss was slightly lower for the PEG300M hybrid; this may be attributed to the higher hydrophilicity and the higher cross-link density of lower molecular weight polymer.

Conclusions

Functionalised PEGM/ 70S30C bioactive hybrids were successfully prepared via sol-gel synthesis. PEGM was successfully used as a cross-linking agent between organic and inorganic components at the molecular level. This resulted in strong covalent Si-O-Si bonds between the organic and inorganic phases which significantly improved the mechanical properties of the resultant hybrids. A corresponding increase in the apatite forming ability was observed for increasing molar fraction of the organic phase; at 50% or below the apatite formation was significantly reduced. Hybrids obtained from higher molecular weight PEGM600 had a higher compressive stress and showed enhanced apatite formation compared to the equivalent PEGM300 hybrid. The 60:40 PEGM600 hybrids were the most promising materials demonstrating good apatite formation, cell proliferation and viability and good mechanical properties. These organic-inorganic hybrid materials exhibited improved mechanical properties and excellent apatite forming ability, confirming their potential use as bone regeneration material.

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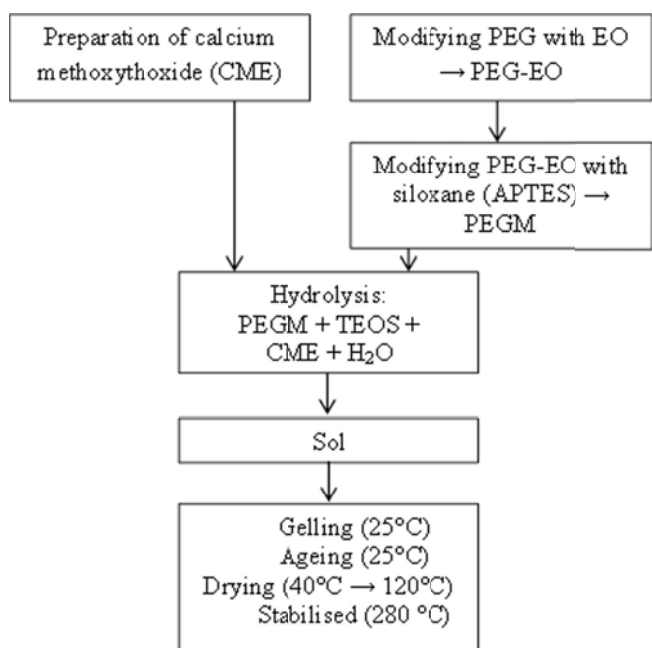


Figure 1. Flow diagram illustrating the hybrid preparation.

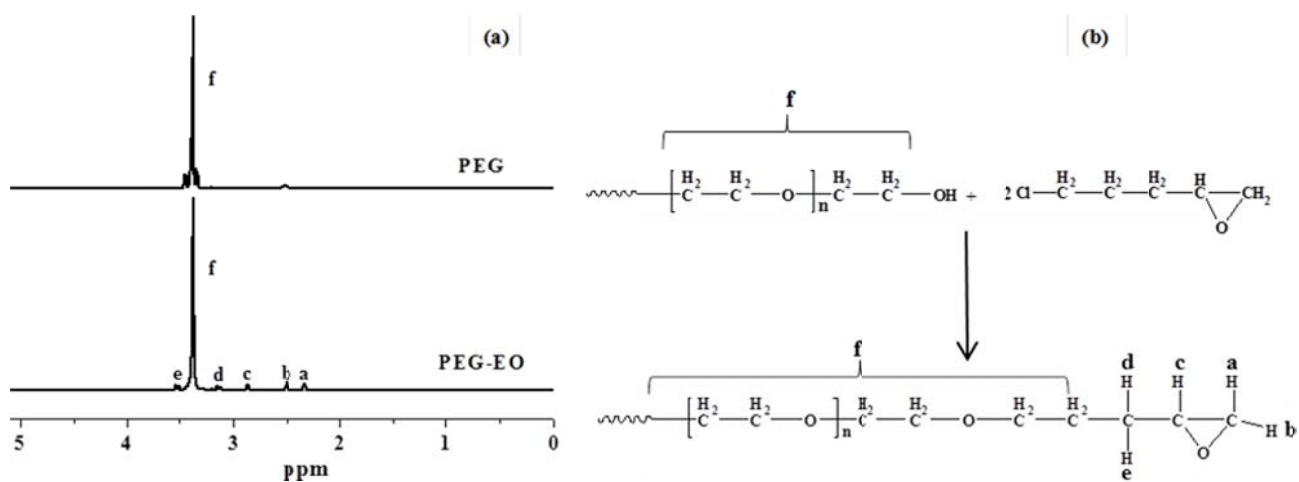


Figure 2(a) ^1H NMR spectra of PEG (top) and the modified PEG-EO polymer (bottom), (b) illustrates the reaction process of PEG to PEG-EO.

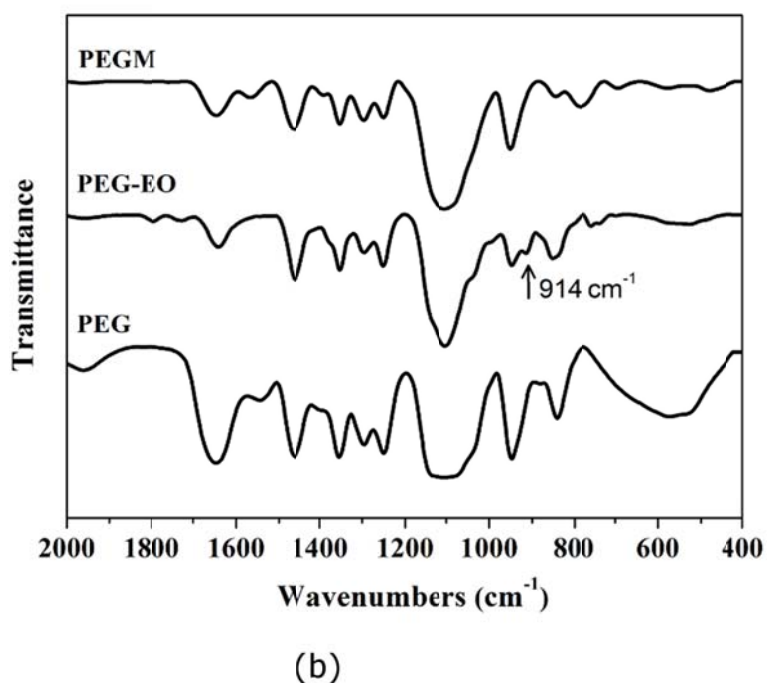
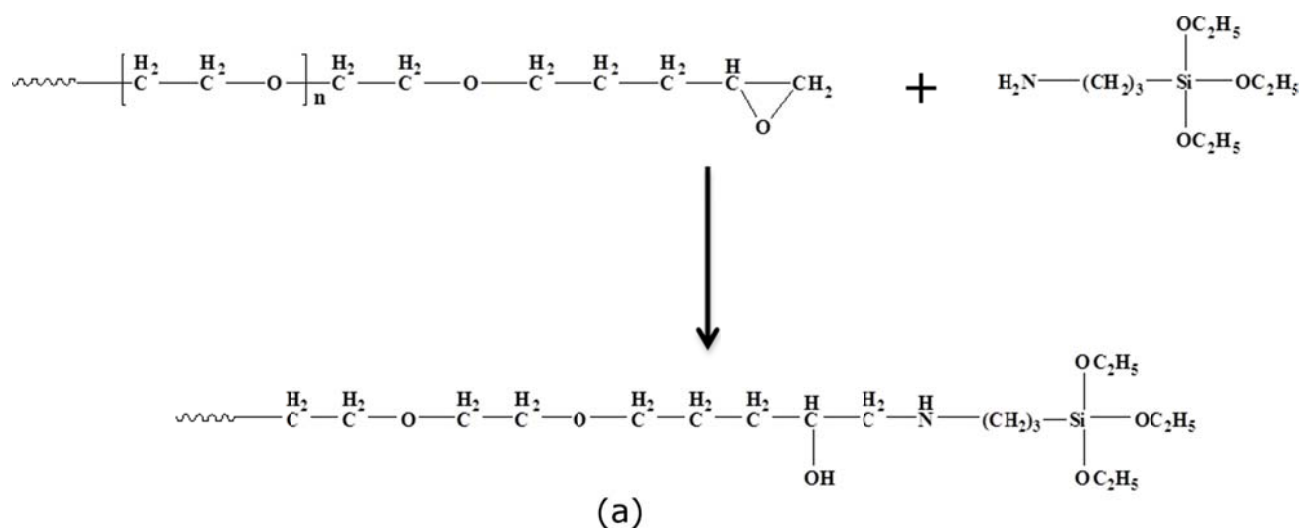


Figure 3. (a) Reaction process illustrating the modification of the PEG-EO end groups, (b) FTIR spectra of PEG, PEG-EO and PEGM. The epoxide group of PEG-EO appears at 914 cm⁻¹.

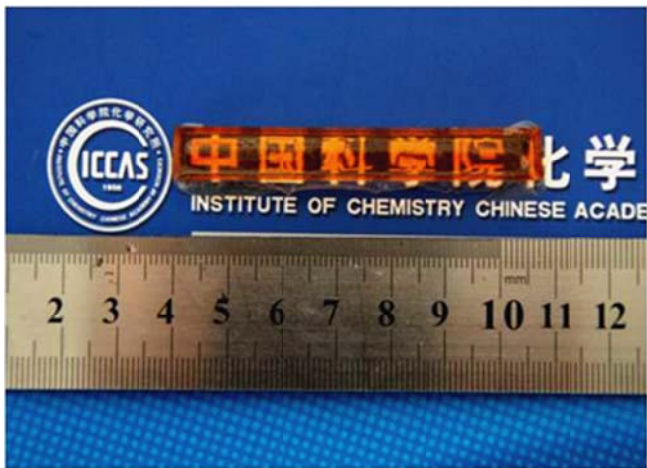


Figure 4. Photo of the 60:40 (organic /inorganic) PEGM600 monolith.

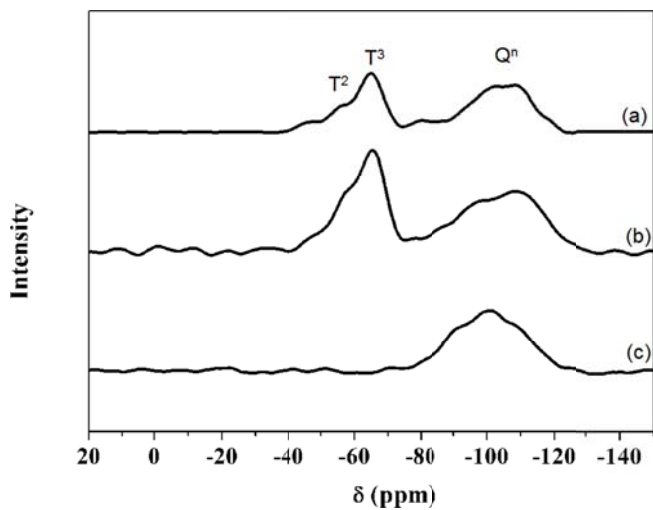


Figure 5. The ^{29}Si solid state MAS-NMR of the sample obtained from different precursors with the ratio of 60/40(organic/inorganic, molar%) : a—PEGM300/TEOS; b—PEGM300/TEOS/CME; c—PEG300/TEOS/CME

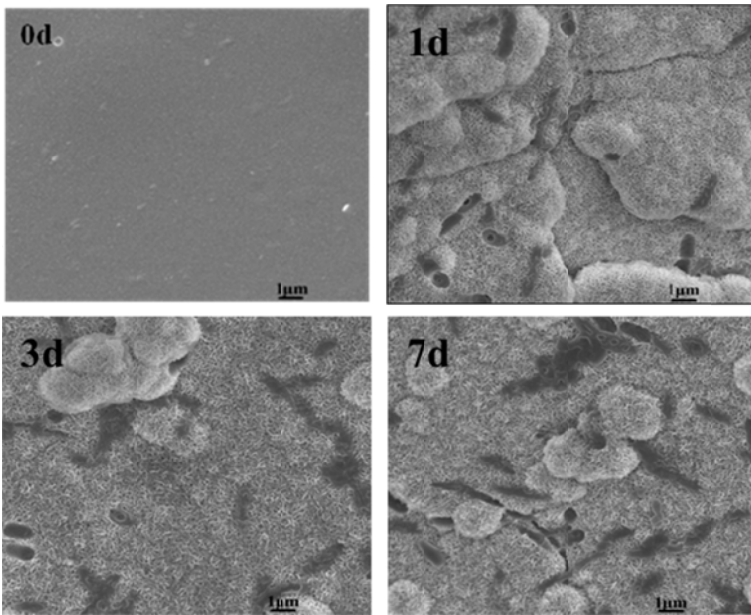


Figure 6. SEM image of hybrid glasses (PEGM600, organic/inorganic=60:40, molar ratio) after immersion in SBF for 0, 1, 3 and 7 days.

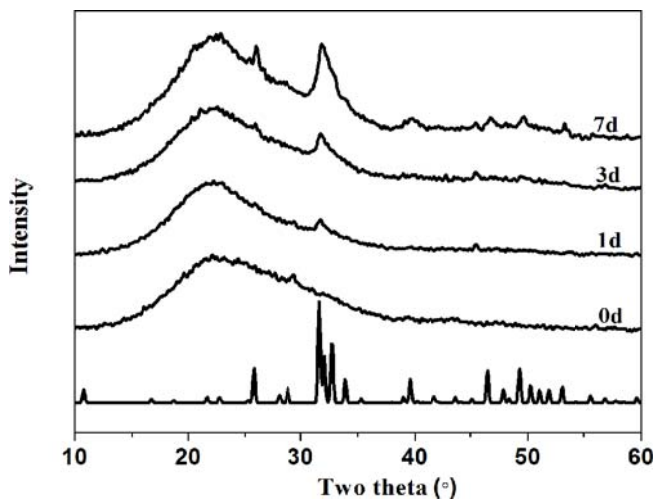


Figure 7. The XRD spectra of hybrid materials (PEGM600, organic/inorganic =60:40, molar ratio) after immersed in SBF for different times.

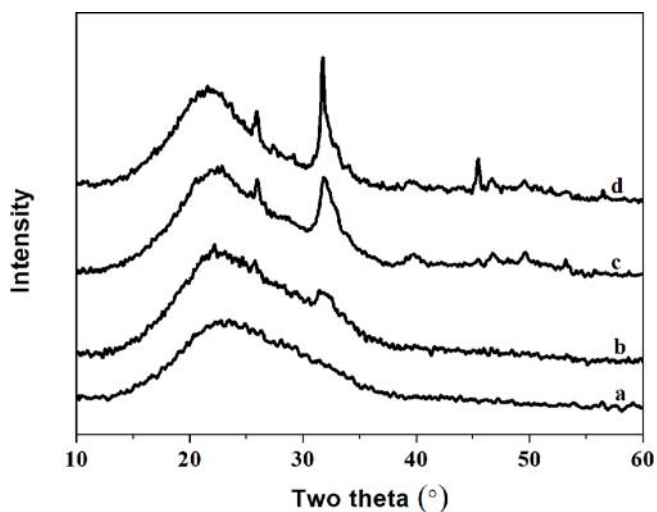


Figure 8. The XRD spectra of hybrid materials with different organic/inorganic (molar ratio, PEGM600) after immersed in SBF for 7 days: a-40:60; b-50:50; c-60:40; d-70:30.

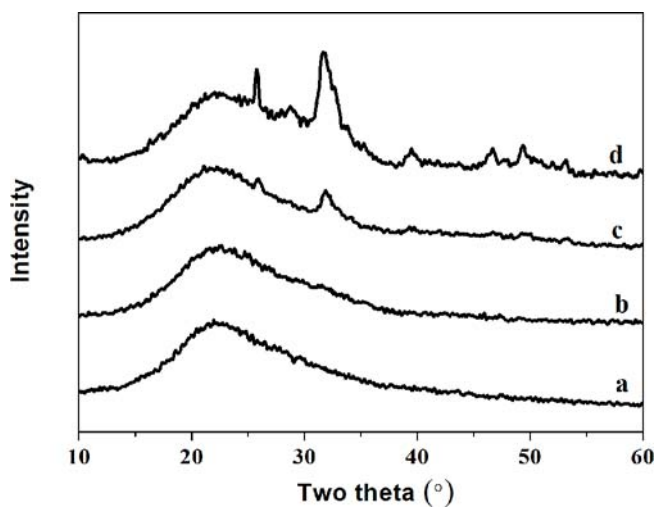


Figure 9. The XRD spectra of hybrid materials with different organic/inorganic (molar ratio, PEGM300) after immersed in SBF for 7 days: a-40:60; b-50:50; c-60:40; d-70:30.

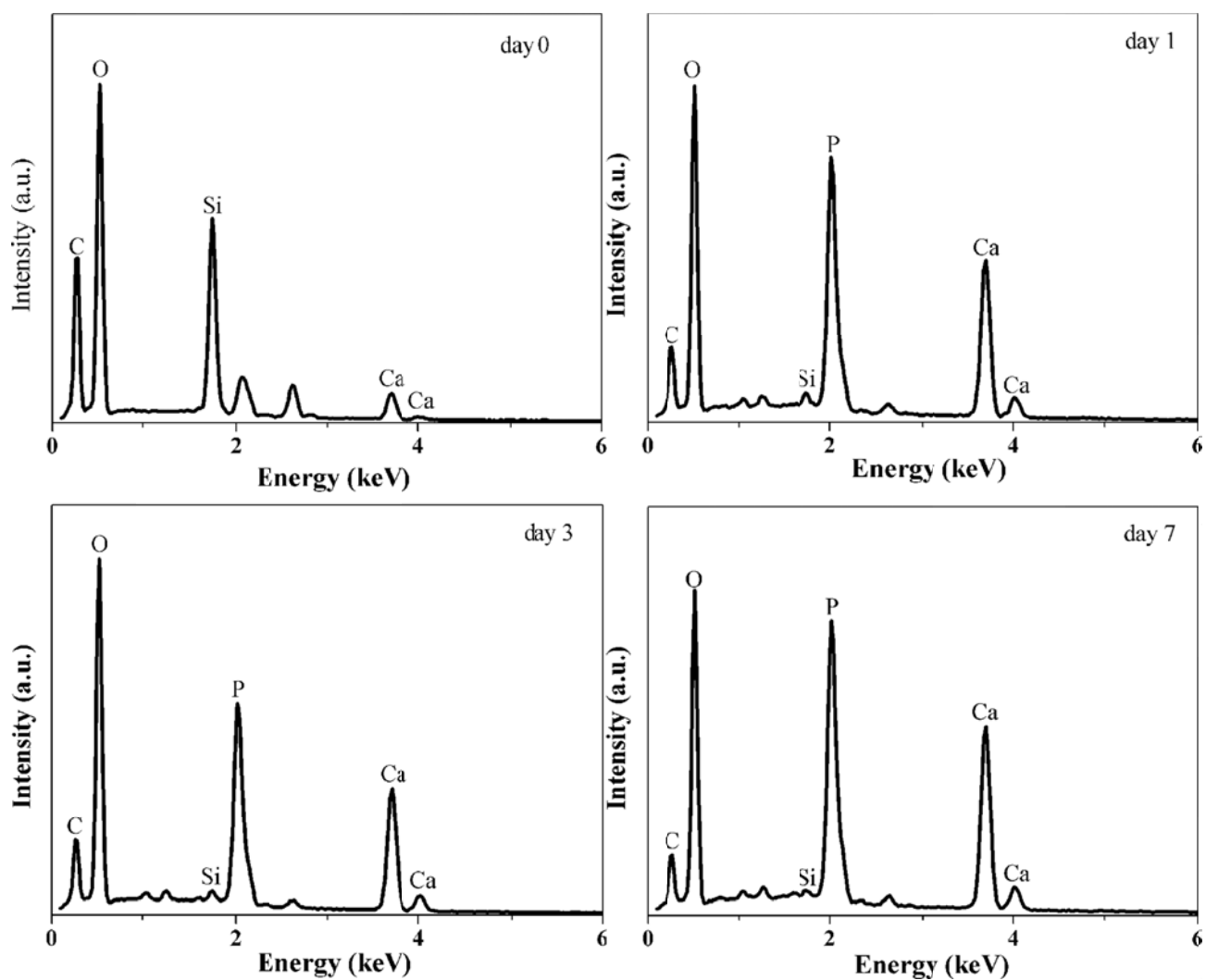


Figure 10. EDXS spectra of the hybrids surface after immersing SBF for 0, 1, 3 and 7 days.

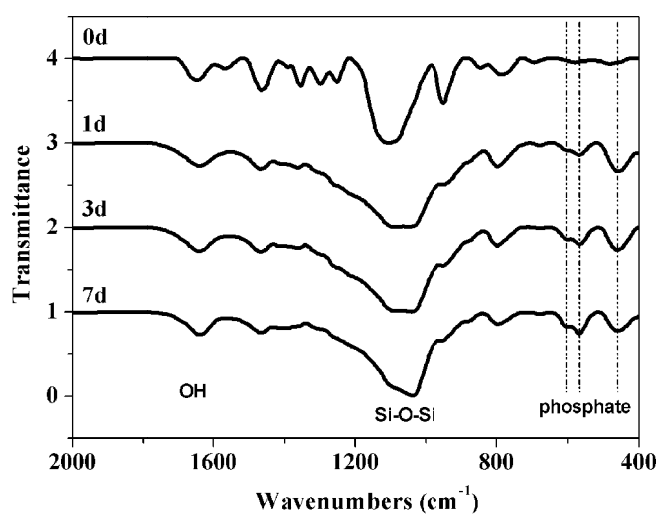


Figure 11. FTIR spectra of the hybrid materials after immersing SBF for 0, 1, 3 and 7 days.

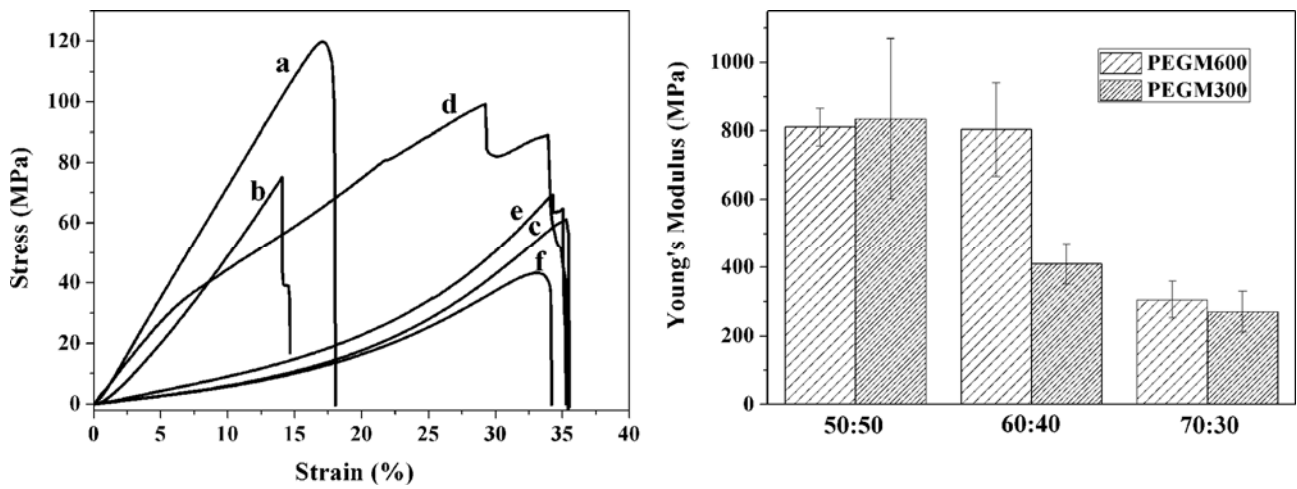


Figure 12. Stress-strain curves (a) and effect of organic/inorganic ratio on Young's Modulus (b) of hybrids obtained by compression test for hybrids with different organic/inorganic ratios (molar%): curve (a), PEGM600 50:50; curve (b), PEGM600 60:40; curve (c), PEGM600 70:30; curve (d), PEGM300 50:50; curve (e), PEGM300 60:40; curve (f), PEGM300 70:30.

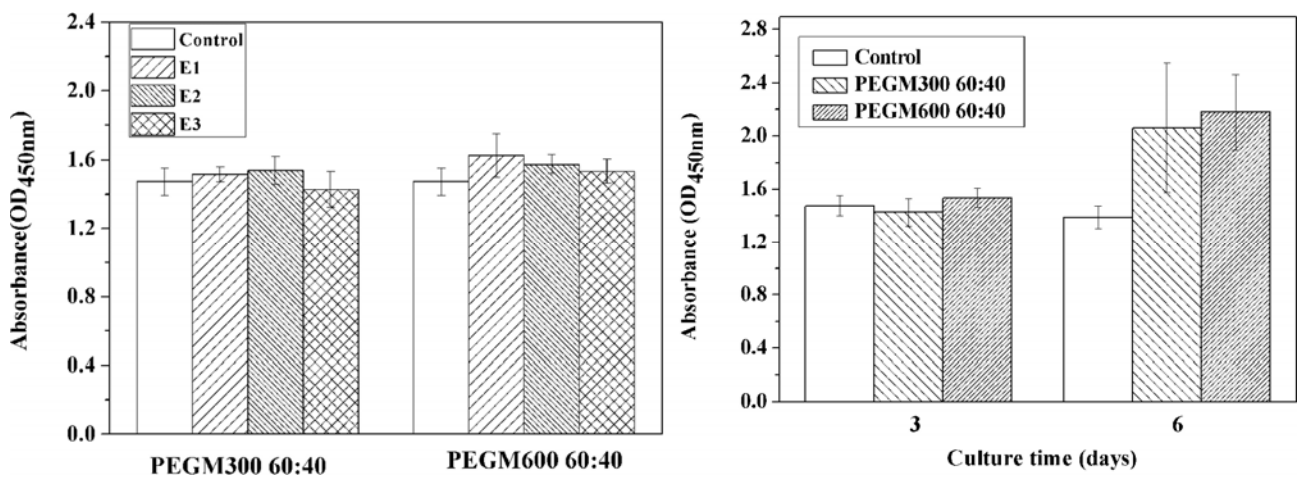


Figure 13. CCK-8 assay for proliferation of MC3T3-E1 cells cultured with (a) extracts (E1, E2 and E3, respectively) of PEGM300 60:40 and PEGM600 60:40 for 3 days and (b) extracts (E3) of PEGM300 60:40 and PEGM600 60:40 for 3 days and 6 days.

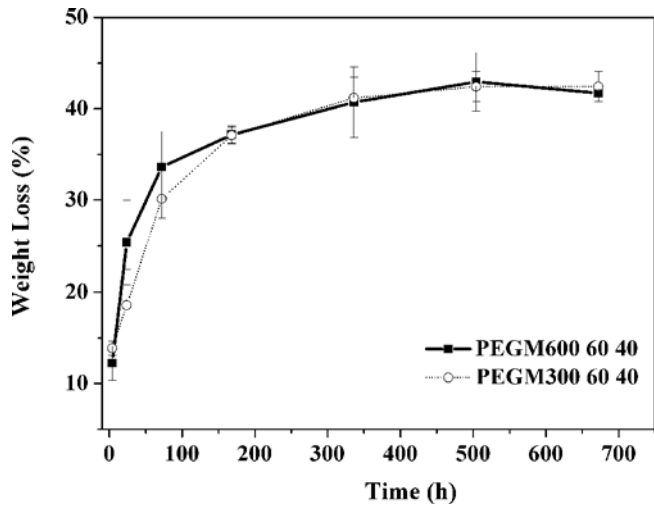


Figure 14. Degradation assay into SBF solution as a function of soaking time for hybrid materials obtained from PEGM600 and PEGM300 with the organic/inorganic ratio of 60:40.

Table 1. Chemical shifts and relative proportions of Tⁿ and Qⁿ species in Class II PEGM/bioactive silica hybrids synthesised (a) without calcium (b) with calcium methoxyethoxide, (c) where unmodified PEG/bioactive silica hybrids. The errors in the chemical shift (δ) are ± 0.5 ppm, in the intensity is $\pm 1\%$ and in the ratios is ± 0.05 .

sample	T ²		T ³		Q ²		Q ³		Q ⁴		T ³ /T _{tot}	Q ⁴ /Q _{tot}
	δ [ppm]	[%]	δ [ppm]	[%]	δ [ppm]	[%]	δ [ppm]	[%]	δ [ppm]	[%]		
a	-59.3	27.3	-65.4	15.6	-	0.0	100.8	16.4	-109.6	40.7	0.4	0.7
b	-61.0	34.3	-66.2	11.0	-	0.0	-101.0	45.1	-112.1	9.6	0.2	0.2
c	-	0.0	-	0.0	-95.2	60.9	100.8	3.9	108.9	35.1	0.0	0.4

Table 2. Compressive strength and Young's modulus values of the hybrids, note it was not possible to measure the unmodified PEG samples which crumbled.

PEGM600			PEGM300		
Organic/inorganic(molar ratio)	Compressive strength(MPa)	Young's Modulus(MPa)	Organic/inorganic(molar ratio)	Compressive strength(MPa)	Young's Modulus(MPa)
50:50	138.1 \pm 36.5	811.2 \pm 53.6	50:50	128.3 \pm 75.1	834.8 \pm 234.4
60:40	81.4 \pm 5.9	804 \pm 136.6	60:40	68.1 \pm 12.6	410.1 \pm 59.6
70:30	52.4 \pm 8.98	305.6 \pm 53.6	70:30	49.17 \pm 15.2	270.1 \pm 60.3
Cancellous bone	4~12	20~500	Compact bone	100~230	3 \times 10 ³ ~3 \times 10 ⁴

Table 3. pH value of samples after immersion in α -MEM

sample	0d	1d	3d	6d
PEGM600 60:40	7.30	7.36	7.41	7.48
PEGM300 60 :40	7.30	7.38	7.49	7.60