β-amyloid (Aβ) deposition in cognitively normal brain, dementia with Lewy bodies, and Alzheimer’s disease: a study using principal components analysis

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Abstract

The densities of diffuse, primitive, and classic β-amyloid (Aβ) deposits were studied in the temporal lobe in cognitively normal brain, dementia with Lewy bodies (DLB), familial Alzheimer’s disease (FAD), and sporadic AD (SAD). Principal components analysis (PCA) was used to determine whether there were distinct differences between groups or whether Aβ pathology was more continuously distributed from group to group. Three principal components (PC) were extracted from the data accounting for 56% of the total variance. Plots of cases in relation to the PC did not result in distinct groups but suggested overlap in Aβ deposition between the groups. In addition, there were linear correlations between the densities of Aβ deposits and the distribution of the cases along the PC in specific brain regions suggesting continuous variation from group to group. PC1 was associated with the degree of maturation of Aβ deposits, PC2 with differences between FAD and SAD, and PC3 with the degree of spread of Aβ pathology into the hippocampus. Apolipoprotein E (APOE) genotype was not associated with variation in Aβ deposition between cases. PCA may be a useful method of studying the pathological interface between closely related neurodegenerative disorders.

Key words: cognitively normal brain, dementia with Lewy bodies (DLB), Alzheimer’s disease (AD), β-amyloid (Aβ) deposits, principal components analysis (PCA).

Introduction

Studies have suggested a significant degree of ‘overlap’ or ‘interface’ between cognitively normal brain, dementia with Lewy bodies (DLB), and Alzheimer’s disease (AD) [6,11,23,24]. Hence, several aspects of AD pathology can be observed in normal aged brain. There is an age-related reduction in brain volume and weight, enlargement of ventricles, and loss of synapses and dendrites in selected areas of normal brain [35]. These changes are accompanied by many of the histological features of AD, viz., senile plaques (SP) and neurofibrillary tangles (NFT) [1,5]. The major molecular constituent of the SP is β-amyloid (Aβ) [23] and hence, Aβ deposition in the form of diffuse (‘pre-amyloid’), primitive (‘neuritic’) [27,28], and classic (‘dense-cored’) deposits is often regarded as a ‘signature’ pathological feature of AD [20,30]. Nevertheless, studies have also demonstrated an overlap in Aβ deposition between AD and normal brain [2,21,40].
Lewy bodies presents with early frontal dementia, cognitive fluctuations, visual hallucinations, syncope, delusions, and rapid eye movement disorder [37]. The clinical and pathological features of DBL may also overlap with AD [24,38] and this potential interface may give rise to difficulties in clinical diagnosis. For example, in a study of 27 clinically diagnosed cases of AD, six were found to have DBL by subsequent pathological examination [43]. An essential feature of the pathological diagnosis of DBL is the presence of α-synuclein-immunoreactive Lewy bodies (LB) in the cerebral cortex and/or brain stem [41]. In addition, many cases of DBL exhibit AD pathology including the presence of SP and NFT [9,22,26,34]. In some DBL cases, the density of SP may be sufficient for a diagnosis of AD using ‘Consortium to Establish a Registry for Alzheimer’s disease’ (CERAD) criteria [42] and these cases are often regarded as examples of ‘mixed’ or ‘multiple pathologies’ [50].

The present study compared the densities of diffuse, primitive, and classic Aβ deposits in regions of the temporal lobe in cognitively normal brain, DBL, familial AD (FAD), and sporadic AD (SAD). The specific objectives were: (1) to determine whether there were distinct differences between groups or whether Aβ pathology was more continuously distributed from group to group and (2) to identify the most important sources of variation in Aβ pathology between patient groups.

Material and methods

Cases

Cases (N = 36, details in Table I) were obtained from the Brain Bank, Department of Neuropathology, Institute of Psychiatry, King’s College, London. Informed consent was given for the removal of all tissue and followed the principles embodied in the 1964 Helsinki declaration (as modified Edinburgh, 2000). Post-mortem delay was less than 20 hours in each case. The control cases (N = 8) had no neurological or psychiatric his-

<table>
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<tr>
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<tr>
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Stories and were matched as closely as possible for age to the AD and DLB cases. The AD cases \((N = 21)\) were clinically assessed and all fulfilled the ‘National Institute of Neurological and Communicative Disorders and Stroke’ and ‘Alzheimer’s Disease and Related Disorders Association’ (NINCDS/ADRDA) criteria for probable AD \([49]\). The histological diagnosis of AD was established by the presence of widespread neocortical senile plaques (SP) consistent with the ‘Consortium to Establish a Registry for Alzheimer’s Disease’ (CERAD) criteria \([42]\). In addition, neurofibrillary tangles (NFT) were abundant in the cerebral cortex and hippocampus \([5]\). Six of the AD cases were familial \([7]\), with two or more generations affected, one of which was linked to a mutation of the presenilin 1 \((PSEN1)\) gene \([45]\). The remaining FAD cases were not associated with mutations of either amyloid precursor protein \((APP)\) or \(PSEN\) genes \([7,19,31,45]\). DLB cases \((N = 7)\) were diagnosed according to the ‘Consortium on Dementia with Lewy bodies’ (CDLB) guidelines \([41]\). Three of these cases had significantly less \(A\beta\) deposits than the others and were diagnosed as ‘pure’ DLB \([22,26,34]\). Apolipoprotein \((APOE)\) genotype, which also influences \(A\beta\) deposition \([16,18,25]\), was determined for 22/36 cases studied.

### Tissue preparation

A block of the temporal lobe, at the level of the lateral geniculate body, was taken from each case and it included the inferior temporal gyrus (ITG), the parahippocampal gyrus (PHG), hippocampus (HC), and dentate gyrus (DG). Tissue was fixed in 10% phosphate buffered formal-saline and embedded in paraffin wax. 7 μm coronal sections were stained with a rab-

<table>
<thead>
<tr>
<th>Case</th>
<th>Group</th>
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<td>83</td>
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</tbody>
</table>

**Table I.** Cont.

DLB – dementia with Lewy bodies, FAD – familial Alzheimer’s disease, SAD – sporadic Alzheimer’s disease, PSEN1 – Presenilin 1, M – Male, F – Female, NA – data not available

* Indicates ‘pure’ DLB with little associated AD pathology, (–) indicates not determined.
bit polyclonal antibody (Gift of Prof. B.H. Anderton, Institute of Psychiatry, King’s College London) raised to the 12-28 amino acid sequence of the Aβ protein [46]. The antibody was used at a dilution of 1 in 1200 and the sections incubated at 4°C overnight. Sections were pretreated with 98% formic acid for 6 minutes which enhances Aβ immunoreactivity. Aβ was visualised using the streptavidin-biotin horseradish peroxidase procedure with diaminobenzidine as the chromogen. Sections were also stained with haematoxylin. The three most common morphological subtypes of Aβ deposit were identified in the sections using previously defined criteria [4,21]. Hence, diffuse deposits were 10-200 μm in diameter, irregular in shape with diffuse boundaries, and lightly stained. Large confluent patches of Aβ immunostaining, which could be a variant of diffuse deposit, were not quantified. Primitive deposits were 20-60 μm, well demarcated, more symmetrical in shape, and strongly stained while classic deposits (Fig. 1B) are well demarcated, more symmetrical in shape, and strongly stained and may be analogous to neuritic plaques, the predominant type of plaque in AD [27,28]. Classic deposits were 20-100 μm, had a distinct central ‘core’ surrounded by a ‘corona’ of dystrophic neurites.

Morphometric methods

In the ITG and PHG, a strip of cortex 17600 to 25600 μm in length, and which included a sulcus and a gyrus, was studied using 1000 × 200 μm contiguous sample fields, the short dimension of the field being aligned with the surface of the pia mater. Hence, the plot included laminae I, II, and most of III, the region containing the highest densities of Aβ deposits in AD [3]. Between 64 and 128 contiguous sample fields were used to sample each gyrus. A micrometer grid with grid lines at intervals of 10 μm was used as the sample field. In the HC, the sample fields were arranged parallel to the alveus to sample sectors CA1, CA2, and CA3. Sampling was then continued into sector CA4 using a guideline marked on the slide and which ceased approximately 400 μm from the DG granule cell layer. In the DG, the lower edge of the sample field was aligned with the top of the granule cell layer as most Aβ deposits were located within the molecular layer. The number of diffuse, primitive, and classic Aβ deposits was counted manually in each field.

Data analysis

Variations in the density and distribution of Aβ deposits in the temporal lobe of the 36 cases were analysed using PCA. PCA measures the degree of similarity between cases (the variables) based on their neuropathological characteristics [10,12,13]. Hence, each case is defined by the density of diffuse, primitive, and classic Aβ deposits in seven regions of the temporal lobe (viz., ITG, PHG, CA1, CA2, CA3, CA4, DG). Preliminary analysis suggested a degree of skew and kurtosis was present indicating a degree of non-normality in the data. Hence, PCA was carried out on the original data and on the data transformed to logarithms [12]. Initially, all PC were extracted which had eigenvalues (λ) > 1 but usually in a PCA, only the first three PC account for significant proportions of the original variance [10]. The result of each PCA is a scatter plot of the 36 cases in relation to the extracted PC in which the distance between cases reflects their degree of similarity or dissimilarity. Hence, if there were distinct differences in Aβ pathology between groups, discrete clusters of cases would be present whereas a more continuous distribution of cases suggests overlapping pathology or a continuum. The PC account for significant proportions of variance in the data, PC1 for the most significant source of variation and PC2 and PC3 for diminishing proportions of the remaining variance. Hence, to identify those aspects of Aβ deposition which may account for this variance, correlations (Pearson’s ‘r’) were calculated between the ‘loadings’ (the coordinates of the case in relation to the PCs) of each case on the PC and Aβ deposit density, age at death, disease duration, and APOE genotype ‘score’ (the sum of the two alleles) in each region.

Results

Examples of diffuse, primitive, and classic Aβ deposits are shown in Fig. 1. Typically, diffuse deposits (Fig. 1A) are irregular in shape with diffuse boundaries, and lightly stained. In contrast, primitive deposits (Fig. 1B) are well demarcated, more symmetrical in shape, and strongly stained while classic deposits (Fig. 1C) have a distinct central ‘core’, usually incorporating one or more neuronal perikarya, and are surrounded by a ‘corona’ of dystrophic neurites. Similar PCA results were obtained using untransformed and transformed data and only the results of the analysis of the original data are reported here. The first three PC extracted from the data accounted for 55.6% of the total variance (PC1 = 27.56%, PC2 = 15.07%, PC3 = 12.97%). The loadings of the cases in relation to PC1 and PC2 and PC3 are shown in Figs. 2 and 3, respectively. No distinct groupings

\[ \beta \text{-amyloid (Aβ) deposition} \]
of cases were evident on either plot suggesting considerable overlap in Aβ deposition between cognitively normal cases, DLB, and AD. In the PC1/2 plot (Fig. 2), the control and DLB cases, especially those designated as ‘pure’ DLB, exhibited high loadings on PC1 but overlapped extensively with the AD cases, which showed a wide distribution over the plot. In addition, the FAD and SAD cases did not appear to cluster in relation to PC1 and PC2, but the FAD cases as a group had consistently lower loadings on PC2 compared with the SAD cases which appear more heterogeneous. FAD cases also exhibited relatively low loadings on PC2 (Fig. 3) including the case linked to a PSEN1 mutation. A proportion of the FAD cases also exhibited high loadings on PC3.

Linear correlations between the extracted PC and the neuropathological variables are shown in Table II. The density of diffuse Aβ deposits in the ITG (r = 0.44, P < 0.01) and PHG (r = 0.36, P < 0.05) were positively correlated with PC1 while the densities of primitive deposits in sector CA3 (r = −0.48, P < 0.01) and classic deposits in sector CA4 (r = −0.48, P < 0.01) were negatively correlated with PC1. The densities of the primitive deposits in the ITG (r = 0.55, P < 0.001) and PHG (r = 0.59, P < 0.001) and classic deposits in the PHG (r = 0.35, P < 0.05) were positively correlated with PC2. The density of the diffuse deposits in sector CA1 was positively correlated with PC3 (r = 0.36, P < 0.05) while the densities of the primitive deposits in sectors CA1 (r = 0.72, P < 0.001) and CA2 (r = 0.58, P < 0.001) and DG (r = 0.62, P < 0.001) were positively correlated with PC3. The density of the classic deposits was negatively correlated with PC3 in the ITG (r = −0.35, P < 0.05) and PHG (r = −0.37, P < 0.01).
positively correlated with PC3 in sector CA2 ($r = 0.43$, $P < 0.01$). In addition, disease duration was positively correlated with PC3 ($r = 0.40$, $P < 0.05$). No significant correlations were observed between the extracted PC and APOE score (PC1: $r = 0.30$, $P > 0.05$; PC2: $r = 0.09$, $P > 0.05$; PC3: $r = -0.05$, $P > 0.05$).

**Discussion**

The objective of this study was to determine whether cognitively normal cases, DLB, FAD, and SAD could be easily distinguished based on the densities of Aβ deposits in the temporal lobe. The PCA suggested: (1) no clear separation of cases into groups and (2) linear correlations in specific regions between the densities of Aβ deposits and the distribution of cases in relation to the PCA. These data suggest continuous change in Aβ density from group to group rather than distinct differences between groups.

Previous studies have reported overlaps in SP or Aβ deposit density between cognitively normal brain and AD. Hence, the distribution of SP was studied in 60 normal elderly cases [39] and it was concluded that it was not possible to distinguish the early stages of AD from normal aging on the basis of SP density alone. Similarly, Bergeron et al. [17] observed SP in 60% of normal elderly cases, albeit at a lower density than in AD. Moreover, Arrigada et al. [14] reported SP in most normal individuals older than 55 years and concluded there may be a ‘continuum’ of pathological change between elderly non-demented brains, early stage AD, and more advanced AD. Aβ deposits are also present in non-demented individuals older than 60 years but are rare before this age [27,40]. In 14 non-demented elderly cases [2], for example, Aβ deposits were present in the temporal lobe in eight cases, with a considerable variation in the density of deposits in control cases and significantly overlapping with AD. By contrast, Gibson [29] studied 119 cases of aging and AD and found amyloid deposits in greater numbers in AD than in normal aging.

The PCA also suggested overlap in Aβ deposition between DLB, cognitively normal brain, and AD. DLB exists in a multiplicity of forms including neocortical, limbic, cerebral, and brainstem types, the neocortical type being the most prevalent [36]. In addition, many cases of DLB exhibit AD pathology including the presence of SP and NFT [22,26,34] with sufficient densi-
ties of SP in some cases for a diagnosis of AD [42]. 
Aβ deposition has also been recorded in DLB [9] and 
may be present at levels similar to ‘pure’ AD. There 
are specific alterations in the ratio of APP isoforms 
common to both DLB and AD suggesting that alternate 
splicing of APP mRNA may play a role in both disorders 
[15] and which could explain this overlap. The presence 
of LB in some cases of AD has blurred further the dis-
tinction between DLB and AD [33,47]. In a family with 
a mutation of the APP gene at codon 717 (APP717), 
one individual had limbic-type DLB, two had neocorti-
tical DLB, while LB were absent in the other family 
members although all exhibited extrapyramidal fea-
tures [43]. To date, 53% of post-mortems of individu-
als with the APP717 mutation have revealed the pres-
ence of LB suggesting a direct link between APP and 
Lewy body formation [43].

PCA identified three axes of variation in Aβ depo-
sition. Hence, PC1 is correlated with variations in den-
sity of diffuse deposits in the ITG and PHG, primitive 
 deposits in sector CA3, and classic deposits in sector 
CA4. Hence, cases with high loadings on PC1, which 
include the majority of cognitively normal, DLB, and 
some AD cases, are characterized by diffuse deposits 
in the ITG and PHG and low densities of primitive and 
classic deposits in sectors CA3/4. Aβ deposit subtypes 
could represent different stages in the maturation of 
a single deposit type [4]. Hence, diffuse deposits may 
represent the earliest stage of Aβ pathology and evolve 
into the primitive and classic Aβ deposits as the dis-
ease progresses, the primitive (‘neuritic’) type deposit 
being the most common in AD [27,28]. Hence, PC1 may 
be associated with the degree of maturation of Aβ 
deposits [4], the density of diffuse deposits being 
greater in control and DLB and more mature deposits 
in AD [27,28].

PC2 is correlated with variations in density of prim-
itive deposits in the ITG and PHG and classic deposits 
in the PHG. Hence, cases with high loadings on PC2, 
which include several SAD, some DLB, and cognitively 
normal cases, are characterized by high densities of 
primitive and classic deposits in the ITG and PHG. By 
contrast, FAD cases exhibit lower densities of primi-
tive and classic deposits in these regions. Hence, PC2 
may be associated largely with variation in Aβ depo-
sition between SAD and FAD cases [7].

PC3 is correlated with variations in density of Aβ 
deposits in sectors CA1/2 of the HC and the DG, cases 
with low loadings on this axis, which include most 
control and DLB cases, having zero or low densities of 
Aβ deposits in these regions. Hence, PC3 is associat-
ed with variation in the spread of Aβ pathology into 
the HC [8]. Disease duration was positively correlated 
with PC3, i.e., longer duration cases were more likely 
to have Aβ deposits in the HC and DG, suggesting that 
this spread may occur later in the disease process. 
In consequence, control and DLB cases are more likely 
ot to have Aβ deposits in these regions although 
deposits may be present in gyri adjacent to the HC.

Table II. Correlations (Pearson’s ‘r’) between 
densities of Aβ deposits in the temporal 
lobe, age at death, disease duration, and apo-
lipoprotein E (APOE) score (sum of the two alle-
les) and the first three principal components (PC)

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<td>0.14</td>
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<td>CA1</td>
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<td>0.72***</td>
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<td>0.08</td>
<td>0.04</td>
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<tr>
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</tr>
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<td>0.43**</td>
</tr>
<tr>
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<td>−0.22</td>
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</tr>
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<td>−0.13</td>
</tr>
<tr>
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<td>0.06</td>
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<td>−0.27</td>
<td>−0.01</td>
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<tr>
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<td>−0.48*</td>
<td>−0.10</td>
<td>0.29</td>
</tr>
<tr>
<td>DG</td>
<td>Diffuse</td>
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<td>−0.27</td>
<td>−0.12</td>
</tr>
<tr>
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<td>Primitive</td>
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<td>0.62***</td>
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<tr>
<td>DG</td>
<td>Classic</td>
<td>0</td>
<td>0.07</td>
<td>−0.07</td>
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ITG = inferior temporal gyrus, PHG = parahippocampal gyrus, CA1-4 = sectors of the hippocampus, DG = dentate gyrus
Significant correlations *P < 0.05, **P < 0.01, ***P < 0.001
APOE genotype was not associated with the PC. APOE genotype has been identified as a major risk factor in AD, individuals with AD having 2-3 times the frequency of allele E4 compared with non-demented elderly controls [48]. Allele E4 appears to accelerate the development of AD pathology within the aged brain and hence, is often associated with an earlier onset of the disease [32]. The relationship between the deposition of Aβ deposits and APOE genotype is controversial but the majority of studies report increased deposition in individuals expressing allele E4 [16,18,25]. Although the present data suggest some increased Aβ deposition in cases expressing APOE allele E4, this source of variation is small compared with that associated with maturation of Aβ deposits, differences between FAD and SAD, and the spread of Aβ deposits into the hippocampus.

In conclusion, changes in the density and distribution of Aβ deposits in the temporal lobe are unlikely to provide a basis for a clear pathological separation of cognitively normal cases, DLB, and AD. Instead, Aβ deposition is more continuously distributed between these groups and cases and three axes of variation were identified: (1) the maturation of diffuse deposits, (2) variation between FAD and SAD, and (3) the spread of Aβ deposits into the HC. APOE genotype was not identified as having a significant influence on Aβ deposition in this study. PCA may be a valuable statistical method of studying pathological changes especially in the interface between closely related neurodegenerative disorders [6,11].

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References


